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Polygenic Prediction using Summary diversion and the second and th

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Polygenic prediction

- Best prediction methods take genetic values as random effect (e.g., BLUP and BayesR).
- These methods require individual-level genotype and phenotype data.
- Data are not publicly accessible, due to privacy and ethical considerations.
- Computationally demanding when numbers of individuals and SNPs are large.
- Use of GWAS summary-level data can address both problems.
- Methodology in human genetics has moved forward to use GWAS summary-level data.



Consensus of sharing GWAS summary data (in human genetics research community)

Has Become a standard to share and make publicly available the summary-level data when publishing a GWAS study.

nature genetics

Asking for more

Because of the usefulness of genome-wide association study (GWAS) data for mapping regulatory variation in the human genome, the journal now asks authors to report the co-location of trait-associated variants with gene regulatory elements identified by epigenetic, functional and conservation criteria. We also ask that authors publish or database the genotype frequencies or association *P* values for all SNPs investigated, whether or not they reached genome-wide significance.



Check for updates

PRIMFR

Cell Genomics



Perspective Workshop proceedings: GWAS summary statistics standards and sharing

2021 S

Jacqueline A.L. MacArthur,^{1,2,*} Annalisa Buniello,¹ Laura W. Harris,¹ James Hayhurst,¹ Aoife McMahon,¹ Elliot Sollis,¹ Maria Cerezo,¹ Peggy Hall,³ Elizabeth Lewis,¹ Patricia L. Whetzel,¹ Orli G. Bahcall,⁴ Inês Barroso,⁵ Robert J. Carroll,⁶ Michael Inouye,^{7,8,9} Teri A. Manolio,³ Stephen S. Rich,¹⁰ Lucia A. Hindorff,³ Ken Wiley,³ and Helen Parkinson^{1,*}

Table 1. Recommended standard reporting elements for GWAS SumStats

Data element	Column header	Mandatory/Optional
variant id	variant_id	One form of variant ID
chromosome	chromosome	is mandatory, either rsID
base pair location	base_pair_ location	or chromosome, base pair location, and genome build ^a
p value	p_value	Mandatory
effect allele	effect_allele	Mandatory
other allele	other_allele	Mandatory
effect allele frequency	effect_allele_ frequency	Mandatory
effect (odds ratio or beta)	odds_ratio or beta	Mandatory
standard error	standard_error	Mandatory
upper confidence interval	ci_upper	Optional
lower confidence interval	ci_lower	Optional

Genome-wide association studies

Emil Uffelmann¹, Qin Qin Huang², Nchangwi Syntia Munung³, Jantina de Vries³, Yukinori Okada^{4,5}, Alicia R. Martin^{6,7,8}, Hilary C. Martin², Tuuli Lappalainen^{9,10,12} and Danielle Posthuma^{1,11}

Table 3 | Databases of GWAS summary statistics

Database	Content	
GWAS Catalog ¹¹⁰	GWAS summary statistics and GWAS lead SNPs reported in GWAS papers	
GeneAtlas ⁸	UK Biobank GWAS summary statistics	
Pan UKBB	UK Biobank GWAS summary statistics	
GWAS Atlas ²⁷³	Collection of publicly available GWAS summary statistics with follow-up in silico analysis	
FinnGen results	GWAS summary statistics released from FinnGen, a project that collected biological samples from many sources in Finland	
dbGAP	Public depository of National Institutes of Health-funded genomics data including GWAS summary statistics	
OpenGWAS database	GWAS summary data sets	
Pheweb.jp	GWAS summary statistics of Biobank Japan and cross-population meta-analyses	

For a comprehensive list of genetic data resources, see REF.¹³. GWAS, genome-wide association studies; SNP, single-nucleotide polymorphism.

CRICOS code 00025B



What do we need to perform summary-databased polygenic prediction?

For simplicity, let's assume the genotypes of each SNP has been standardised with column mean zero and variance one when conducting GWAS.

In this case, the minimum data required are

- SNP marginal effect estimates
- GWAS sample size
- LD correlations among SNPs



SNP marginal effect estimates

GWAS estimates effect of each SNP one at a time from single SNP regression, so the estimate is marginal to (unconditional on) other SNPs.

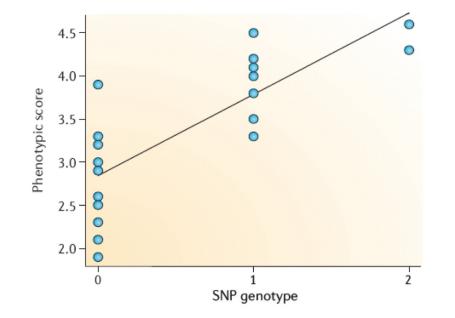
 $b_j = \left(\mathbf{X}_j'\mathbf{X}_j\right)^{-1}\mathbf{X}_j'\mathbf{y}$

Assuming **X** has been standardised with column mean zero and variance one, then

$$\mathbf{X}_{j}'\mathbf{X}_{j} = nVar(\mathbf{X}_{j}) = n$$

And

$$b_j = \frac{1}{n} \mathbf{X}'_j \mathbf{y}$$



Note that it has the inner product of the SNP genotypes and the phenotypes.



SNP marginal effect estimates

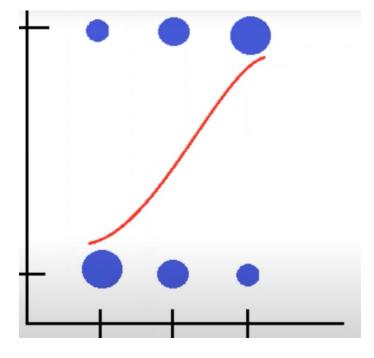
For diseases, GWAS is done using logistic regression

$$\log\left(\frac{p_i}{1-p_i}\right) = \mu + X_{ij}b_j$$

The SNP effect is log odds ratio (OR), i.e., difference in log odds for cases vs. controls

$$b_j = \log(OR)$$

Approximately equal to the b_j from the linear model when true effect size is small.



0 1 2 Genotype



What do we need to perform summary-databased polygenic prediction?

For simplicity, let's assume the genotypes of each SNP has been standardised with column mean zero and variance one when conducting GWAS.

Then, the minimum data required are

- SNP marginal effect estimates
- GWAS sample size
- LD correlations among SNPs

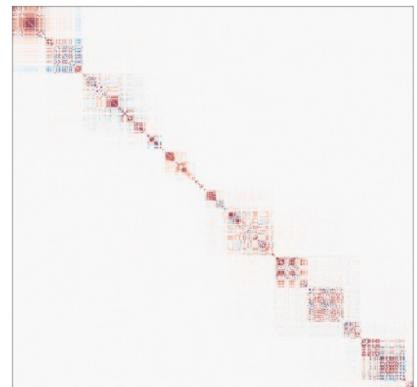


Linkage disequilibrium (LD) correlations

Usually obtained from a reference population LD correlation matrix

$$\mathbf{R} = \frac{1}{n} \mathbf{X}' \mathbf{X}$$

assuming **X** is standardised with mean zero and variance one





Use of summary data only - how does it work?

GWAS results and LD correlations are **sufficient statistics** for the estimation of SNP joint effects!



A statistic is **sufficient** if no other statistic that can be calculated from the same sample provides any additional information as to the value of the parameter.

e.g., $x_1, x_2, ..., x_n \sim N(\mu, \sigma^2)$ and we want to estimate μ and σ^2

$$\hat{\mu} = \frac{\sum_{i=1}^{n} x_i}{n}$$
 • $\sum_{i=1}^{n} x_i$ and *n* are sufficient statistics for μ

$$\hat{\sigma}^2 = \frac{\sum_{i=1}^n x_i^2}{n} - \left[\frac{\sum_{i=1}^n x_i}{n}\right]^2 \qquad \bullet \quad \sum_{i=1}^n x_i^2 \text{, } \sum_{i=1}^n x_i \text{ and } n \text{ are sufficient} \\ \text{statistics for } \sigma^2$$

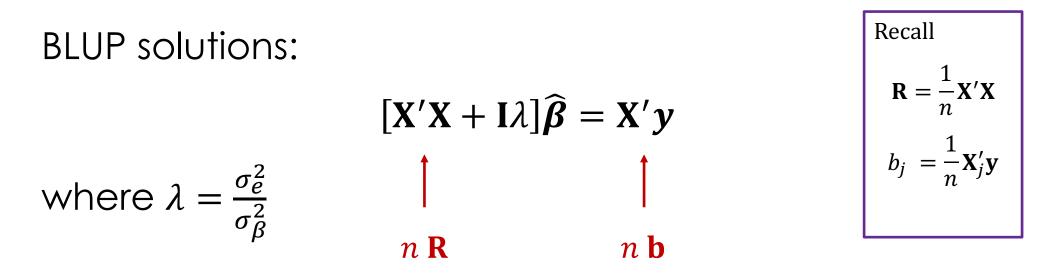
We don't need to know the value of each x!

How does it work?



BLUP

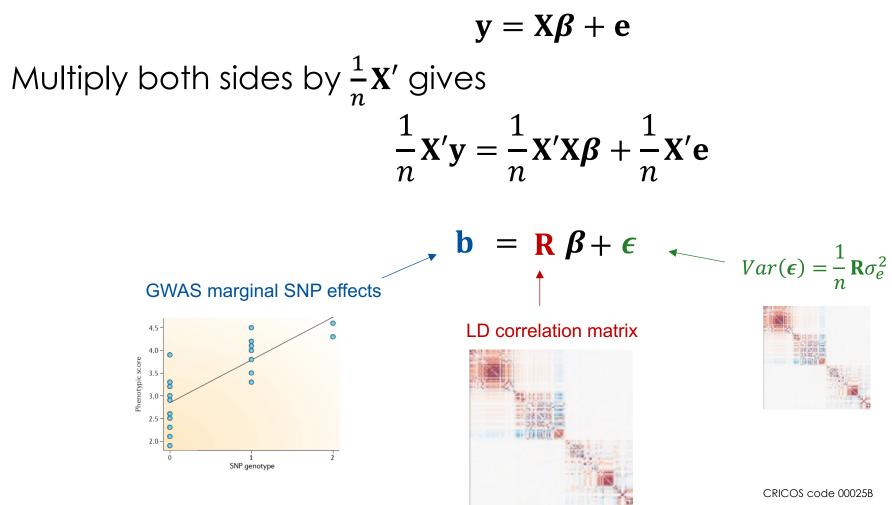




R (LD matrix), **b** (marginal effects) and n are sufficient statistics for the estimation of β .



Consider an individual-data model with a standardised genotype matrix **X**:



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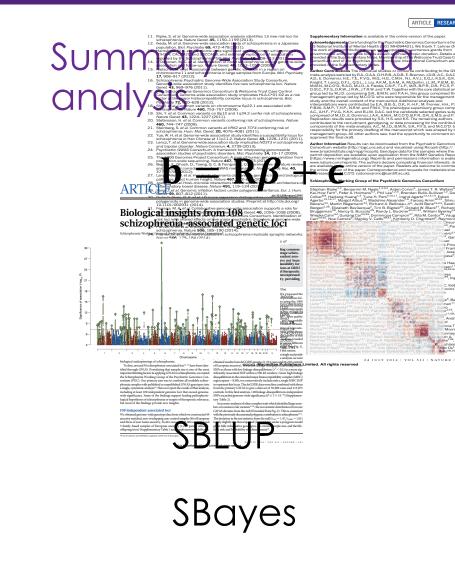
Individual-level data analysis

$$\mathbf{y} = \mathbf{X}\boldsymbol{\beta} + \mathbf{e}$$



BLUP

Bayes



Covariates, such as age and sex, are accounted for when conducting GWAS

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BLUP vs. SBLUP

BLUP

- $y = X\beta + e$
- $Var(\mathbf{e}) = \mathbf{I}\sigma_e^2$
- $[\mathbf{X}'\mathbf{X} + \mathbf{I}\lambda]\widehat{\boldsymbol{\beta}} = \mathbf{X}'\mathbf{y}$
- $\hat{\beta} = [X'X + I\lambda]^{-1}X'y$ Genotype matrix Phenotypes matrix Individual-level data Summary-level data

SBLUP

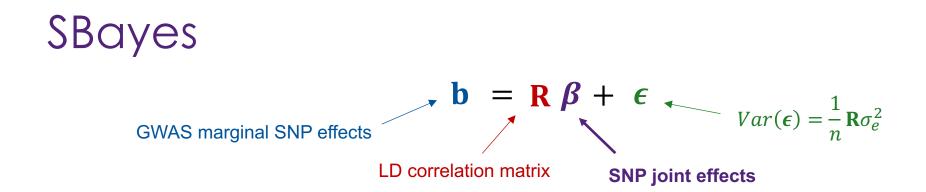
- **b**= $\mathbf{R}\boldsymbol{\beta} + \boldsymbol{\epsilon}$
- $Var(\boldsymbol{\epsilon}) = \frac{1}{n} \mathbf{R} \sigma_e^2$
- $[n\mathbf{R} + \mathbf{I}\lambda]\widehat{\boldsymbol{\beta}} = n\mathbf{b}$
- $\widehat{\boldsymbol{\beta}} = [n\mathbf{R} + \mathbf{I}\lambda]^{-1}n\mathbf{b}$

LD

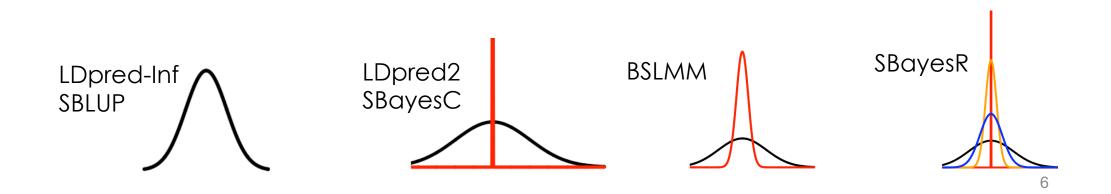
correlation

matrix





Prior distribution for each SNP effect



SBayesR

Each SNP effect has a mixture distribution:

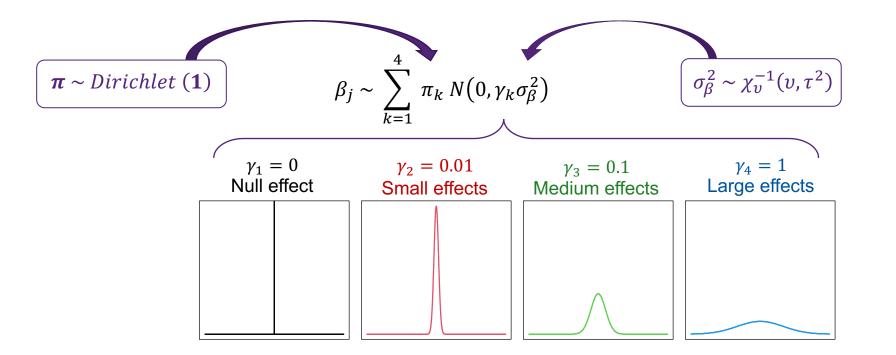


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https://doi.org/10.1038/s41467-019-12653-0 OPEN

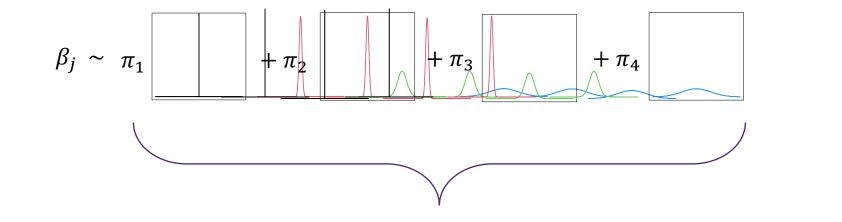
Improved polygenic prediction by Bayesian multiple regression on summary statistics

Luke R. Lloyd-Jones ^{1,9*}, Jian Zeng ^{1,9*}, Julia Sidorenko^{1,2}, Loïc Yengo¹, Gerhard Moser^{3,4}, Kathryn E. Kemper¹, Huanwei Wang ¹, Zhili Zheng¹, Reedik Magi², Tõnu Esko², Andres Metspalu^{2,5}, Naomi R. Wray ^{1,6}, Michael E. Goddard⁷, Jian Yang ^{1,8*} & Peter M. Visscher ^{1*}





Account for various SNP effect distributions



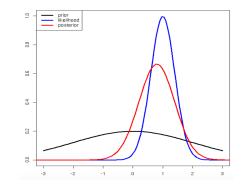




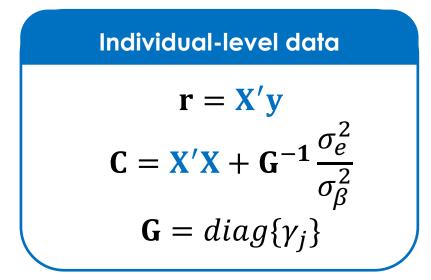
The posterior distribution of SNP effects

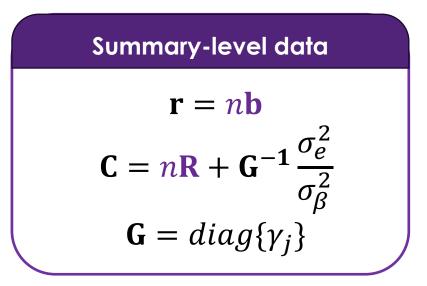
Posterior \propto Likelihood \times Prior $f(\beta | \text{Summary data}) \propto f(\text{Summary data} | \beta) \times f(\beta)$

 $\boldsymbol{\beta} | \mathbf{b} \sim \mathrm{N}(\mathbf{C}^{-1}\mathbf{r}, \mathbf{C}^{-1}\sigma_e^2)$



where



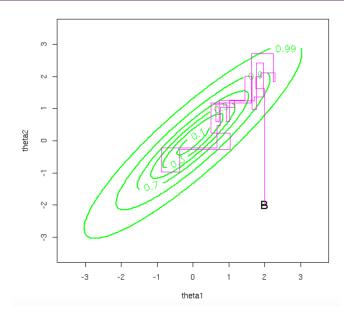




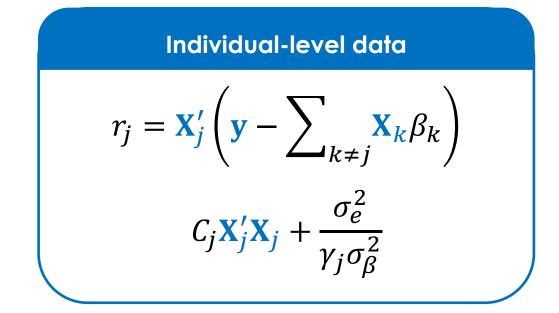
Single-site Gibbs sampling

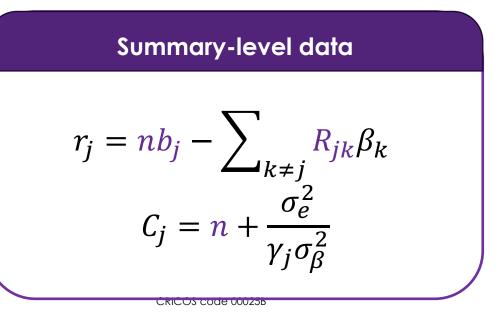
Full conditional distribution for β_j

$$f(\beta_j \mid \mathbf{b}, else) = N\left(\frac{r_j}{C_j}, \frac{\sigma_e^2}{C_j}\right)$$



where







Comparison between individual and summary level algorithms

Algorithm 1 – Individual level data algorithm

Initialise parameters and read genotypes and phenotypes in PLINK binary format Initialise $\mathbf{y}^* = \mathbf{y} - \mathbf{X}\boldsymbol{\beta}$ for i :=1 to number of iterations do **for** i :=1 **to** *p* **do** Calculate $r_i^* = \mathbf{x}_i' \mathbf{y}^*$ Calculate $r_i = r_i^* + \mathbf{x}_i' \mathbf{x}_i \beta_i^{(i-1)}$ Calculate $\sigma_c^2 = \sigma_\beta^2 \gamma_{\delta_i=c}$ for each of *C* classes (e.g., BayesR C=4 and $\gamma = (0, 0.0001, 0.001, 0.01)$) Calculate the left hand side $l_{jc} = \mathbf{x}'_{j}\mathbf{x}_{j} + \frac{\sigma_{c}^{2}}{\sigma^{2}}$ for each of the *C* classes Calculate the log densities of given $\delta_j = c$ using $\log(\mathcal{L}_c) = -\frac{1}{2} \left[\log \left(\frac{\sigma_c^2 l_{jc}}{\sigma_c^2} \right) - \frac{r_j^2}{\sigma_c^2 l_{jc}} \right] + \log(\pi_c)$, where π_c is the current Calculate the full conditional posterior probability for $\delta_j = c$ for *C* classes with $\mathbb{P}(\delta_j = c | \boldsymbol{\theta}, \mathbf{y}) = \frac{1}{\sum_{l=1}^{C} \exp[\log(\mathcal{L}_l) - \log(\mathcal{L}_l)]}$ Using full conditional posterior probabilities sample class membership for $\beta_i^{(i)}$ using categorical random variable sampler Given class sample SNP effect $\beta_i^{(i)}$ from $N\left(\frac{r_j}{L_i}, \frac{\sigma_e^2}{L_i}\right)$ Given SNP effect adjust corrected phenotype side $(\mathbf{y}^*)^{(i)} = (\mathbf{y}^*)^{(i-1)} - \mathbf{x}_i \left(\beta_i^{(i)} - \beta_i^{(i-1)} \right)$ od Sample update from full conditional for σ_{β}^2 from scaled inverse chi-squared distribution $\tilde{v}_{\beta} = v_{\beta} + q$ and $\tilde{S}_{\beta}^2 = \frac{v_{\beta}S_{\beta}^2 + \sum_{j=1}^{q} \frac{v_j}{\gamma_c}}{v_j}$ where *q* is the number of non-zero variants Sample update from full conditional for σ_e^2 from scaled inverse chi-squared distribution $\tilde{\nu}_e = n + \nu_e$ and scale parameter $\hat{S}_{\ell}^2 = \frac{SSE + v_c \hat{S}_{\ell}^2}{n + v_c}$ and $\hat{SSE} = \mathbf{y}^* \mathbf{y}^*$ Sample update from full conditional for $\boldsymbol{\pi}$, which is Dirichlet($C, \mathbf{c} + \boldsymbol{\alpha}$), where \mathbf{c} is a vector of length C and contains the counts of the number of variants in each variance class and $\alpha = (1, ..., 1)$ Calculate genetic variance for h_{SNP}^2 calculation using $\sigma_g^2 = \text{Var}(\mathbf{X}\boldsymbol{\beta})$ Calculate $h_{SNP}^2 = \frac{\sigma_{\tilde{g}}}{\sigma_2^2 + \sigma_3^2}$ od

Algorithm 2 Summary data algorithm

Initialise parameters and read summary statistics Reconstruct X'X and X'y from summary statistics and LD reference panel Calculate $\mathbf{r}^* = \mathbf{X}'\mathbf{v} - \mathbf{X}'\mathbf{X}\boldsymbol{\beta}$ for i :=1 to number of iterations do **for** i :=1 **to** *p* **do** Calculate $\mathbf{r}_i = \mathbf{r}_i^* + \frac{\mathbf{x}_i'\mathbf{x}_i}{\beta}$ Calculate $\sigma_c^2 = \sigma_{\alpha}^2 \gamma_{\delta_i=c}$ for each fo *C* classes (e.g., SBayesR C=4 and $\gamma = (0, 0.01, 0.1, 1)'$) Calculate the left hand side $l_{jc} = \frac{\mathbf{x}'_{j}\mathbf{x}_{j}}{\sigma_{c}^{2}}$ for each of the *C* classes Calculate the log densities of given $\delta_j = c$ using $\log(\mathcal{L}_c) = -\frac{1}{2} \left[\log \left(\frac{\sigma_c^2 l_{jc}}{\sigma_c^2} \right) - \frac{r_j^2}{\sigma_c^2 l_{jc}} \right] + \log(\pi_c)$, where π_c is the current Calculate the full conditional posterior probability for $\delta_j = c$ for C classes with $\mathbb{P}(\delta_j = c | \boldsymbol{\theta}, \mathbf{y}) = \frac{1}{\sum_{\ell=1}^{C} \exp[\log(\mathcal{L}_l) - \log(\mathcal{L}_c)]}$ Using full conditional posterior probabilities sample class membership for $\beta_i^{(i)}$ using categorical random variable sampler Given class sample SNP effect $\beta_i^{(i)}$ from $N\left(\frac{\mathbf{r}_i}{L_i}, \frac{\sigma_e^2}{L_i}\right)$ Given SNP effect adjust corrected right hand side $(\mathbf{r}^*)^{(i+1)} = (\mathbf{r}^*)^{(i)} - \mathbf{X'}\mathbf{x}_i \left(\beta_i^{(i+1)} - \beta_i^{(i)}\right)$. $\mathbf{X'}\mathbf{x}_i$ is the *j*th column of $\mathbf{X'}\mathbf{X}$. od Sample update from full conditional for σ_{α}^2 from scaled inverse chi-squared distribution $\tilde{\nu}_{\alpha} = \nu_0 + q$ and $\tilde{\tau}_{\alpha}^2 = \frac{\nu_0 \tau_0^2 + \Sigma_{j=1}^q \frac{P_j}{\tau_{\delta_j}^2}}{\nu_{\alpha+q}}$ where *q* is the number of non-zero variants Sample update from full conditional for σ_e^2 from scaled inverse chi-squared distribution $\tilde{\nu}_e = n + \nu_e$ and scale parameter $\tilde{\tau}_{c}^{2} = \frac{SSE + v_{c}\tau_{c}^{2}}{n + v_{c}}$ and $SSE = \mathbf{y}'\mathbf{y} - \boldsymbol{\beta}'\mathbf{r}^{*} - \boldsymbol{\beta}'\mathbf{X}'\mathbf{y}$ Sample update from full conditional for $\boldsymbol{\pi}$, which is Dirichlet($C, \mathbf{c} + \boldsymbol{\alpha}$), where \mathbf{c} is a vector of length C and contains the counts of the number of variants in each variance class Calculate genetic variance for h_{SNP}^2 calculation using $\sigma_g^2 = MSS/n$, where $MSS = \hat{\beta}' \mathbf{X}' \mathbf{y} - \hat{\beta}' r^*$ Calculate $h_{SNP}^2 = \frac{\sigma_g^2}{\sigma_q^2 + \sigma_g^2}$ od

X'y and X'X can be replaced by *n*b and *n*R

Lloyd-Jones & Zeng et al. 2019 NC supplement



Relax the assumption of standardised genotypes

- Insofar, derivations are based on standardised genotypes.
- GWAS are usually performed using unstandardised genotypes (allele counts; X_i^*)
- b_i^* from the GWAS using X_i^* is called per-allele effect
- Need to rescale GWAS effects by $b_j = s_j b_j^*$ where s_j is the genotype SD
- Because

$$y = X_j^* b_j^* + e$$
$$= \frac{X_j^*}{s_j} \times s_j b_j^* + e$$
$$= X_j b_j + e$$



How to find s_j

We need more information and make assumptions!

Method 1:

- Use (minor) allele frequency p_j and assume Hardy-Weinberg Equilibrium (HWE)
- $s_j = \sqrt{2p_j(1-p_j)}$
- Cons: allele frequency from the GWAS sample may not be available

Method 2:

• Use GWAS effect standard error SE_j and assume b_j contribute to negligible variance

•
$$SE_j = \sqrt{\frac{1}{\mathbf{X}_j' \mathbf{X}_j} \hat{\sigma}_e^2} = \sqrt{\frac{1}{ns_j^2} \left(\sigma_y^2 - s_j^2 b_j^2\right)} \rightarrow s_j = \sqrt{\frac{\sigma_y^2}{nSE_j^2 + b_j^2}} \approx \sqrt{\frac{\sigma_y^2}{nSE_j^2}}$$

• Cons: b_j may be large for a major QTL.



How to find s_j

In GCTB, we use

Method 3:

- Use p_j and SE_j and estimate σ_y^2
- σ_y^2 = the median value of $2p_j(1-p_j)[nSE_j^2+b_j^2]$ across SNPs

•
$$s_j = \sqrt{\frac{\sigma_y^2}{nSE_j^2 + b_j^2}}$$

• The median value of per-SNP σ_y^2 is robust to allele frequency errors.



Critical information from GWAS summary data

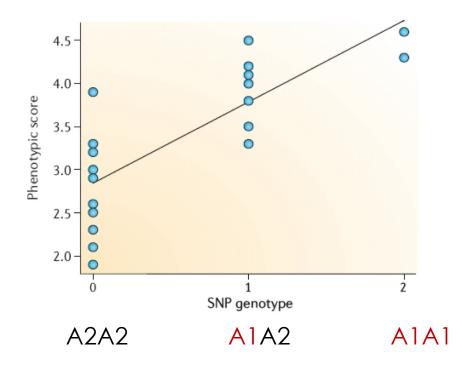
- Marginal SNP effects
- GWAS sample size
- Standard errors
- Allele frequencies



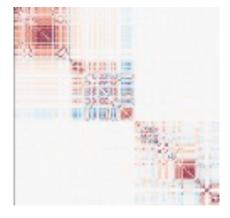
Other information critical to quality control (QC)

Which allele is the effect allele in GWAS?

e.g., A1 allele



Need to match with the allele used to calculate the LD matrix in the reference sample





Other information critical to quality control (QC)

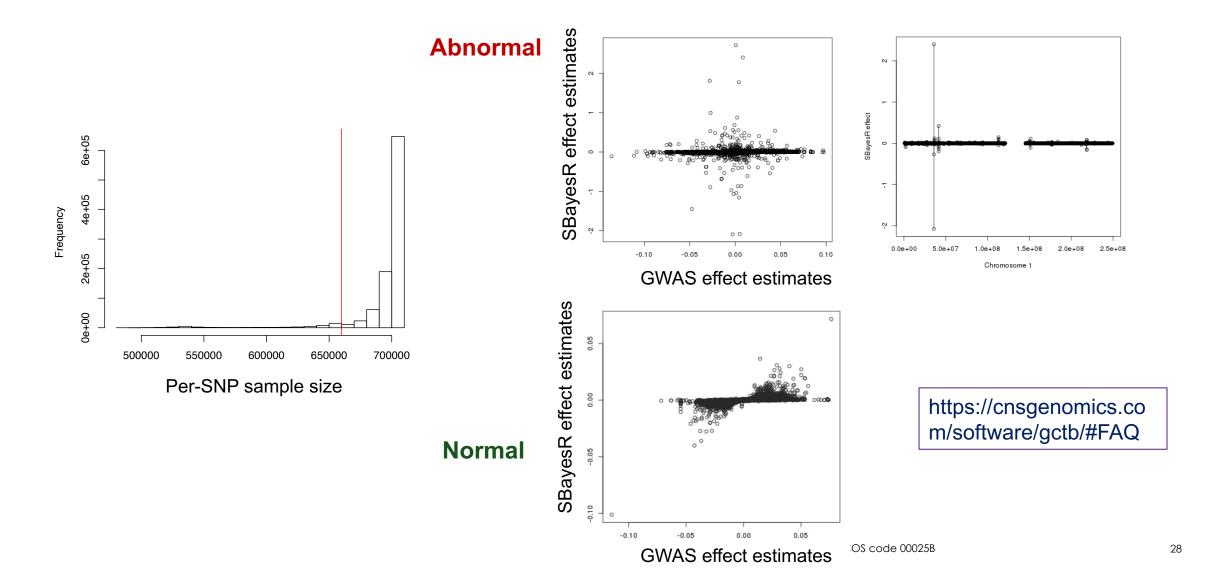
Per-SNP sample size

Heterogeneity in per-SNP sample size (usually due to meta-analysis) may result in a convergence problem in MCMC.

We recommend to visualise the per-SNP sample size distribution and remove the outliers.



Influence of heterogeneity in per-SNP sample size





Critical information from GWAS summary data

- Marginal SNP effects
- (Per-SNP) GWAS sample sizes
- Standard errors
- Effect alleles and alternate alleles (A1 and A2)
- Effect allele frequencies

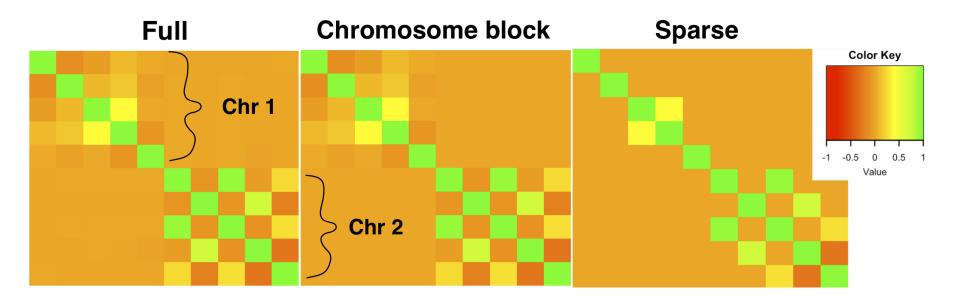
Input file (.ma)

SNP A1 A2 freq b se p N
rs1001 A G 0.8493 0.0024 0.0055 0.6653 129850
rs1002 C G 0.0306 0.0034 0.0115 0.7659 129799
rs1003 A C 0.5128 0.0045 0.0038 0.2319 129830



LD matrix from a reference sample

- Often cannot use genome-wide full LD matrix from the GWAS sample.
- Use a reduced (banded, sparse, or shrunk) LD matrix from a reference sample.



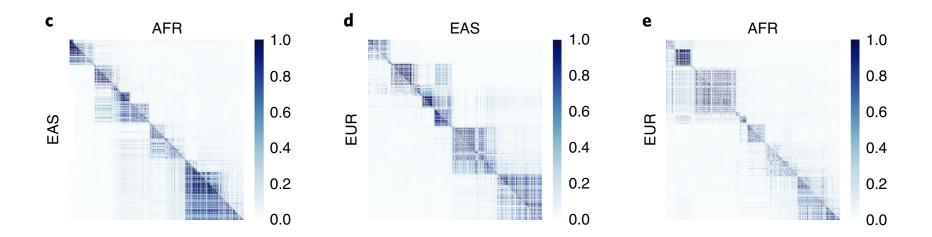


LD re to • No systematic atterences in LD → s

tches with GWAS population in genetics

• No systematic atterences in LD \rightarrow same ancestry and population structure

Minimu mplin rianc
 LD → LD ref sample size cannot be too small





Implicit assumptions

LD reference population matches with GWAS population in genetics

- No systematic differences in LD \rightarrow same ancestry and population structure
- Minimum sampling variance in LD \rightarrow LD ref sample size cannot be too small

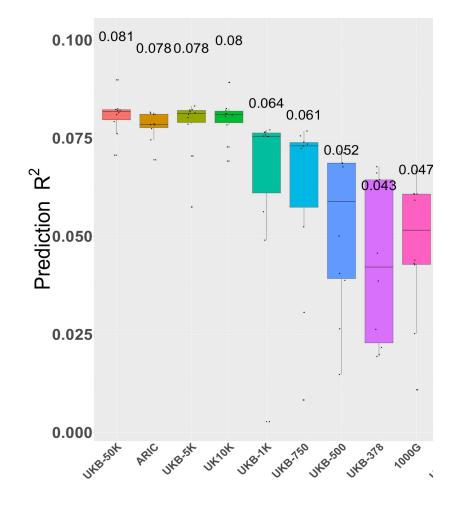
GWAS data are collected on the same set of individuals

- Often an issue in GWAS meta-analysis
- Consistent genotyping platforms or imputation panels across cohorts
- Remove SNP outliers in per-SNP sample size

Violation these assumptions can cause model misspecification, resulting in attenuated prediction accuracy or even failure to reach convergence.



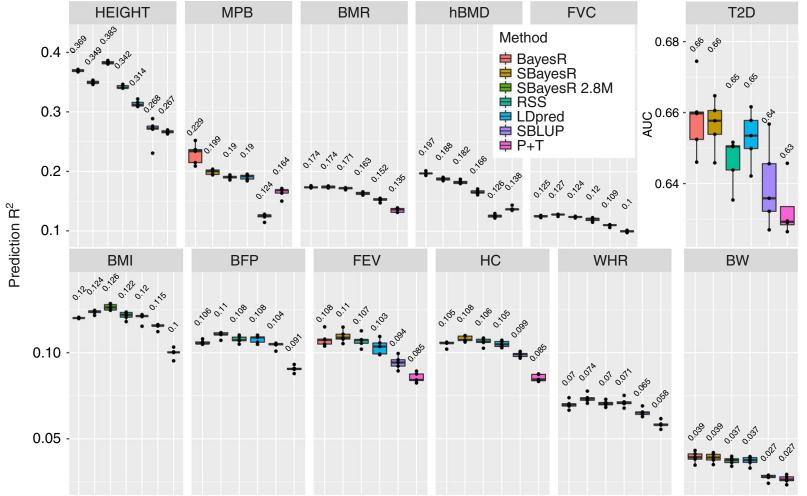
Influence of choice of LD reference



Lloyd-Jones & Zeng et al. 2019 NC



Method comparison



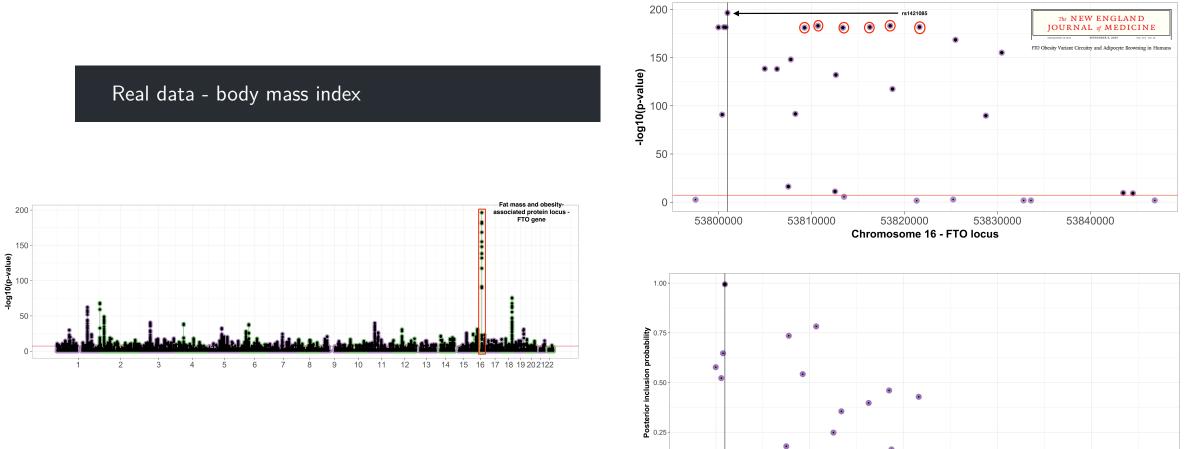
Lloyd-Jones & Zeng et al. 2019 NC

CRICOS code 00025B



Real data - FTO - GWAS

Fine-mapping



•

53800000

0.00 -

CRICOS code 00025B

53820000

Chromosome 16 - FTO locus

53810000

•

• •

53840000

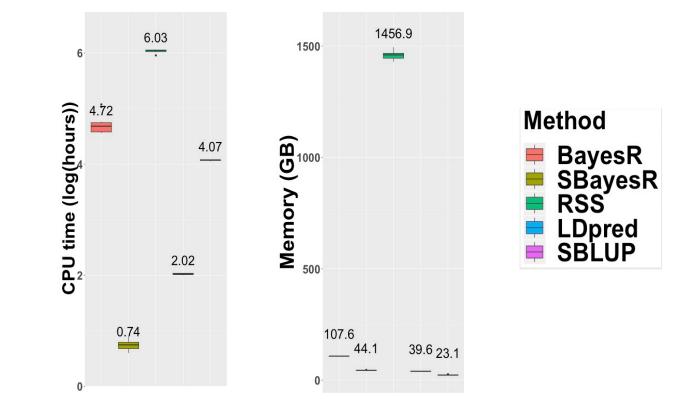
53830000



Computational efficiency

100k individuals 1M SNPs

Resource consumption for summary-data-based methods is independent of sample size once GWAS summary statistics are obtained.





Summary

- Summary-level methods unleash the full power of GWAS of large sample sizes for polygenic prediction.
- Free from limitation of data accessibility.
- Computationally efficient.
- Only an approximation to the individual-level counterpart due to reduction in LD matrix.
- Flexible to incorporate other information, such as functional annotations or omics data.



Practical 5: Polygenic prediction using summary data

https://cnsgenomics.com/data/teaching/GNGWS23/model5/Practical5_SBLUP_and_SBayes.html

Log into the cluster

cd to your working directory in scratch: cd /scratch/[your folder]

You will learn how to run MCMC with summary data using the toy example data set in R.

You will use GCTB to run SBayesR in the simulated data set based on real genotypes. Now we are able to leverage information from the full UK Biobank data without accessing to the individual genotypes and phenotypes.