

Connectivity Map for identifying drug candidates

Lecture Overview

- Gene signature matching
- A database of compound gene signatures - CMap
- Generating a disease gene signature
- Querying CMap

GWAS to medicine



- Are GWAS-significant genes targets of existing drugs (identify drug repurposing candidates)
 - Repurposing FDA-approved compounds – better safety profile, lower risk, shortest path to approval
 - Screening failed drugs against new indications - benefit-risk profile may vary depending on the unmet medical need
- But...
 - Drugs with unknown mechanism of action (MoA) will be missed with this approach
 - Important disease biology may be lost under stringent p-value thresholds

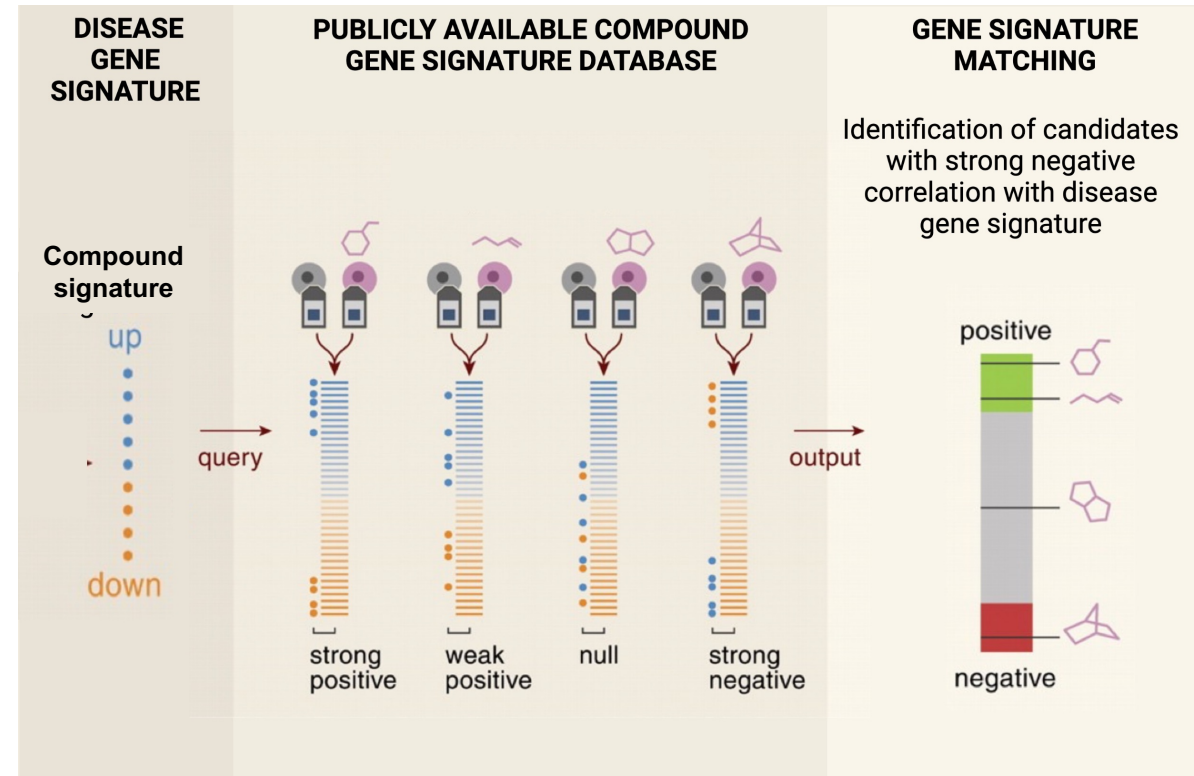
Gene signature matching

Gene expression signature matching

Assumption: compounds that have the same MoA induce similar gene expression responses. Can be useful for:

1. Understanding MoA of a compound
2. Drug repurposing potential
3. Identifying new drug candidates
 - Compounds that reverse gene expression changes associated with disease
 - Does not require knowledge of the drug's MoA
4. Identifying potential drug side-effects

Requires gene expression signatures for drugs and diseases



Connectivity Map (CMap)

Library of gene expression signatures in response to chemical and genetic perturbation.

- >1 million gene expression profiles
- ~50 different cell lines (only 4 are non-cancer cell lines)
- ~20,000 compounds (chemical perturbation)
- ~20,000 knockdown/overexpression (genetic perturbations)

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The Connectivity Map: Using Gene-Expression Signatures to Connect Small Molecules, Genes, and Disease

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<https://www.broadinstitute.org/connectivity-map-cmap>

1st Generation CMap - Lamb et al Science 2013

- Need to establish the relation among diseases, physiological processes, and the action of small-molecule therapeutics.
- Previous compound and genetic perturbation studies in yeast and rats
 - Translation to humans
 - High cost of animal studies
- Mammalian cells
 - Generalisable, systematic and biologically relevant
 - BUT...a large number of parameters would need to be optimized for each perturbation – cell type, dose, duration
- Pilot study demonstrated the feasibility of this approach

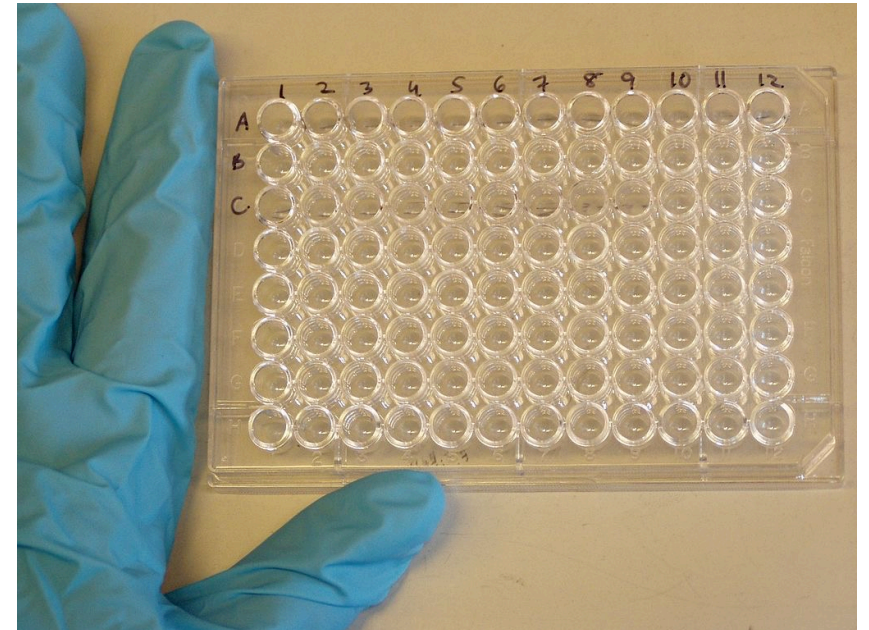
1st Generation CMap - compounds

164 distinct small-molecule perturbagens, selected to represent a broad range of activities:

- FDA–approved drugs
- nondrug bioactive “tool” compounds
- multiple compounds sharing molecular targets (test if they share gene signatures e.g. HDAC inhibitors)
- compounds with the same clinical indication (test whether compounds with different MoA that treat the same disease generate similar gene signatures e.g. antidiabetics)
- Molecules that are proximal (e.g. selective estrogen receptor modulators) and distal to gene expression
- Molecules whose targets are not expressed in the cell types being tested (COX2 inhibitors)

1st Generation CMAP – cell lines

- Stably grown over long periods of time
- breast cancer epithelial cell line MCF7
 - extensively molecularly characterised,
 - used as a reference cell line
 - amenable to culture in 96-well plates
- prostate cancer epithelial cell line PC3
- nonepithelial lines HL60 (leukemia) and SKMEL5 (melanoma)
- Context-dependent gene signatures



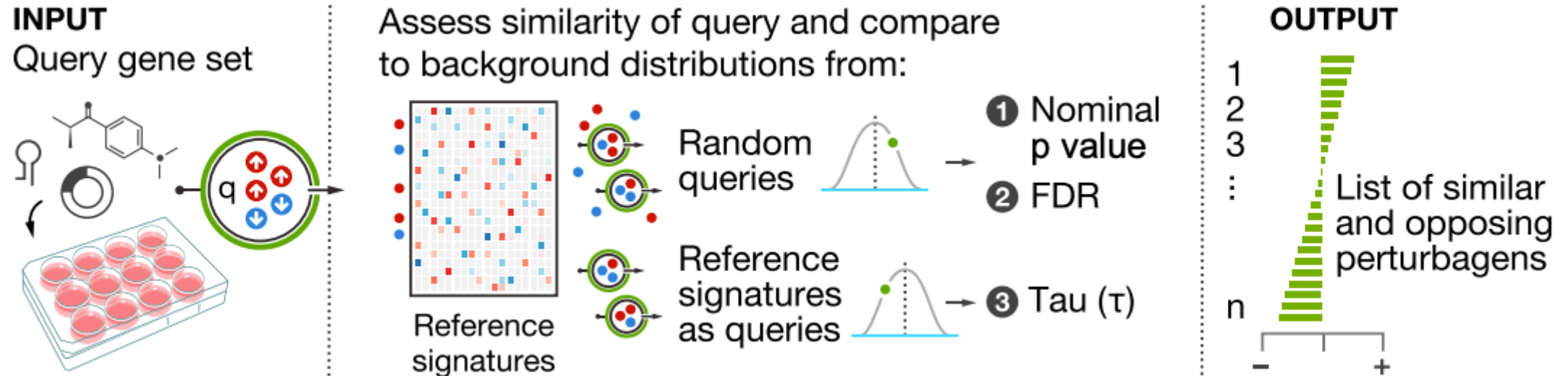
1st Generation CMAP – dose and duration

- 10uM – optimal concentration is not known for many compounds
 - Toxicity studies required for proper optimisation of dose
- 6 and 12 hrs post-treatment
 - Profiles obtained too early might not yield robust signals—esp for perturbations that do not directly modulate transcription
 - Profiles obtained too late may reflect secondary and tertiary responses
 - obtain signatures related to direct mechanisms of action
- Dose and duration dependent on question of interest, but difficult to optimise in such high-throughput experiments.

Compound gene signature generation

- Control perturbations for each treatment (cells grown on the same plate treated with vehicle only)
 - minimize the impact of batch-to-batch
 - biological and technical variation
- Replicates
- Data were collected in multiple batches over a period of 1 year by Affymetrix GeneChip microarrays.
- DEG analysis – compound-treated gene expression vs intra-batch vehicle-treated control
- For each treatment ~22,000 genes rank-ordered according to differential expression

Connectivity score



- Used non-parametric, rank-based pattern-matching strategy based on the Kolmogorov-Smirnov statistic (GSEA).
- Tau score - fraction of reference gene sets with a greater similarity to the perturbagen than the current query.

Example results – HDAC inhibitors

- HDACs – remove acetyl groups on histones and regulate gene expression
- Determine if a query signature can recover compounds from the same class (same MoA).
- Query derived from response of bladder and breast cancer cells treated with 3 HDAC inhibitors (vorinostat, MS-27-275, trichostatin)
 - 13-gene (8 up and 5 down-regulated) signature
- Off-target effects

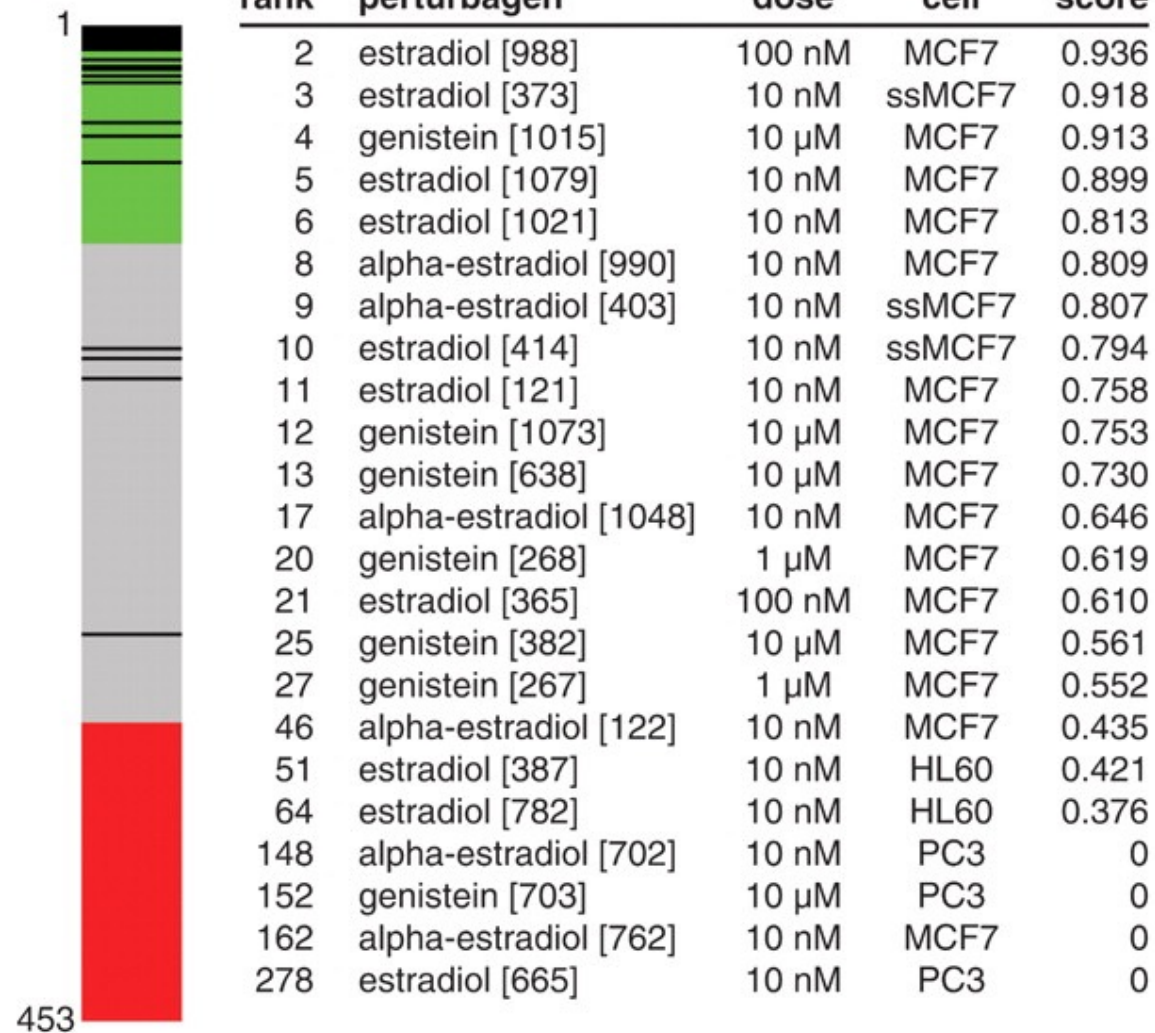
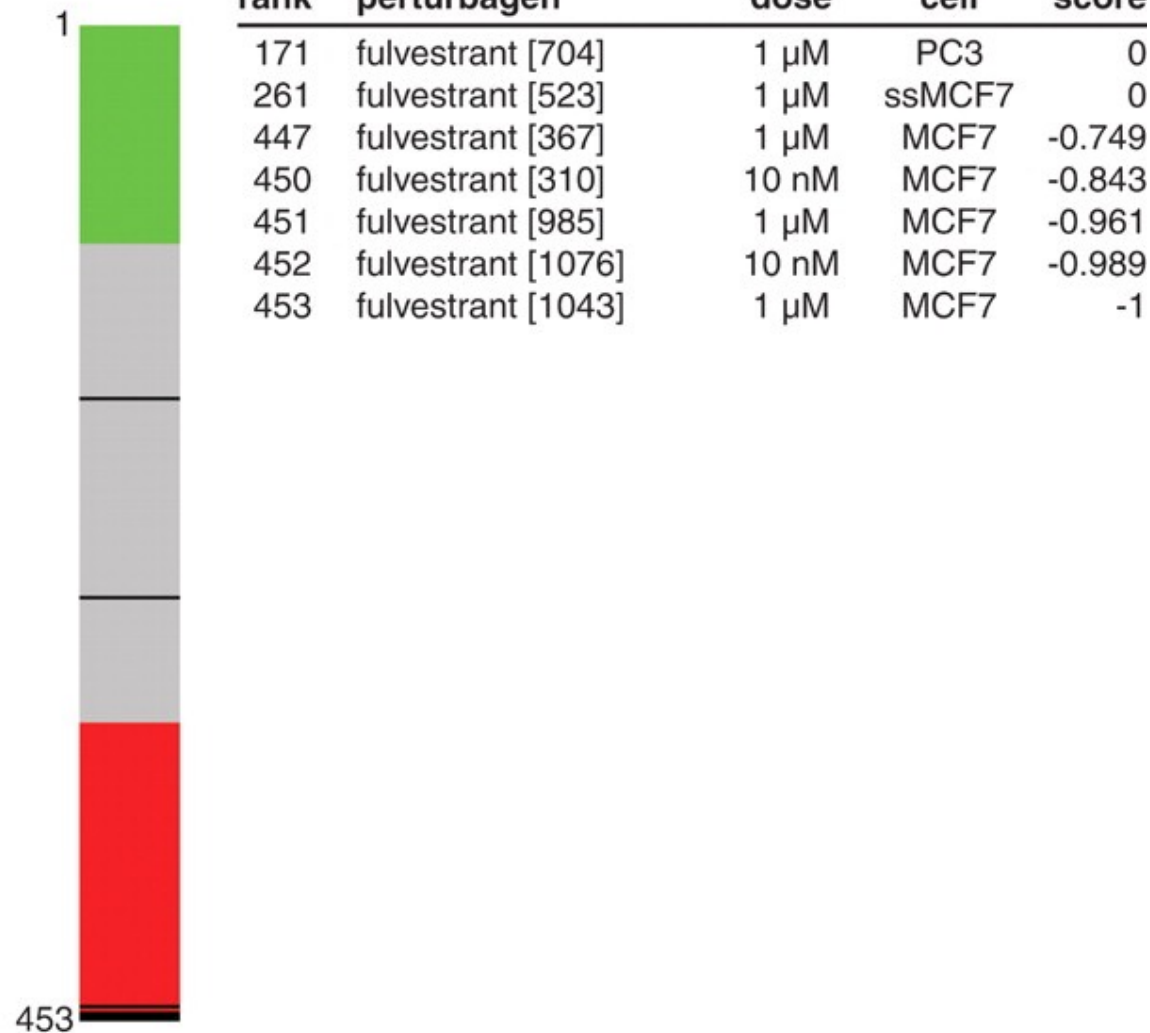
A



rank	perturbagen	dose	cell	score
1	vorinostat [1000]	10 μ M	MCF7	1
2	trichostatin A [873]	1 μ M	MCF7	0.969
3	trichostatin A [992]	100 nM	MCF7	0.931
4	trichostatin A [1050]	100 nM	MCF7	0.929
5	vorinostat [1058]	10 μ M	MCF7	0.917
6	trichostatin A [981]	1 μ M	MCF7	0.915
7	HC toxin [909]	100 nM	MCF7	0.914
8	trichostatin A [1112]	100 nM	MCF7	0.908
9	trichostatin A [1072]	1 μ M	MCF7	0.906
10	trichostatin A [1014]	1 μ M	MCF7	0.893
11	trichostatin A [332]	100 nM	MCF7	0.882
12	trichostatin A [331]	100 nM	MCF7	0.846
13	trichostatin A [448]	100 nM	PC3	0.788
14	valproic acid [345]	10 mM	MCF7	0.743
15	valproic acid [23]	1 mM	MCF7	0.735
16	valproic acid [1047]	1 mM	MCF7	0.733
17	trichostatin A [413]	100 nM	ssMCF7	0.725
18	valproic acid [410]	10 mM	HL60	0.725
19	valproic acid [458]	1 mM	PC3	0.680
33	valproic acid [409]	1 mM	HL60	0.634
39	valproic acid [1020]	500 μ M	MCF7	0.619
52	valproic acid [346]	2 mM	MCF7	0.582
61	valproic acid [1078]	500 μ M	MCF7	0.563
71	valproic acid [629]	1 mM	SKMEL5	0.539
72	valproic acid [347]	500 μ M	MCF7	0.539
73	valproic acid [989]	1 mM	MCF7	0.538
76	valproic acid [433]	1 mM	PC3	0.528
89	trichostatin A [364]	100 nM	HL60	0.507
92	valproic acid [497]	1 mM	ssMCF7	0.501
297	valproic acid [348]	50 μ M	MCF7	0
388	valproic acid [994]	200 μ M	MCF7	0
403	valproic acid [1002]	50 μ M	MCF7	0
419	valproic acid [1060]	50 μ M	MCF7	-0.537

Example - Estrogens

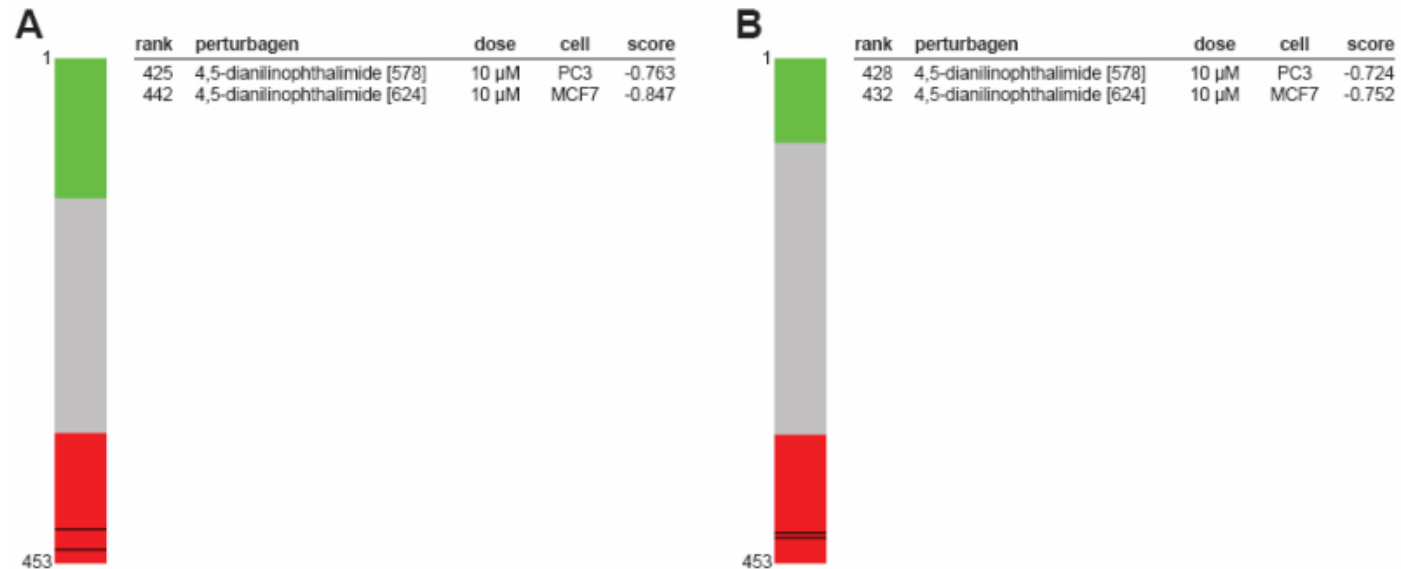
- Estrogen – modulates nuclear hormone signaling by binding to estrogen receptor.
- Query signature – MCF7 cells treated with 17beta-estradiol
 - 129-gene signature (40 up and 89 down-regulated)

A

B


Connections with Disease States

- Query – DEGs from a rat model of diet-induced obesity
- Several differences in exp design: Rat vs human, exposure duration – 65 days vs 6 hrs, adipose tissue vs cell lines

Fig. S4. PPAR γ Agonists are Connected with Diet-induced Obesity in Rats. Barview (as Fig. 2) showing all instances of troglitazone ($n=2$), rosiglitazone ($n=1$), indometacin ($n=1$) and 15-delta prostaglandin J2 ($n=1$) in PC3 cells. Unabridged results from this query are provided as Result S8.



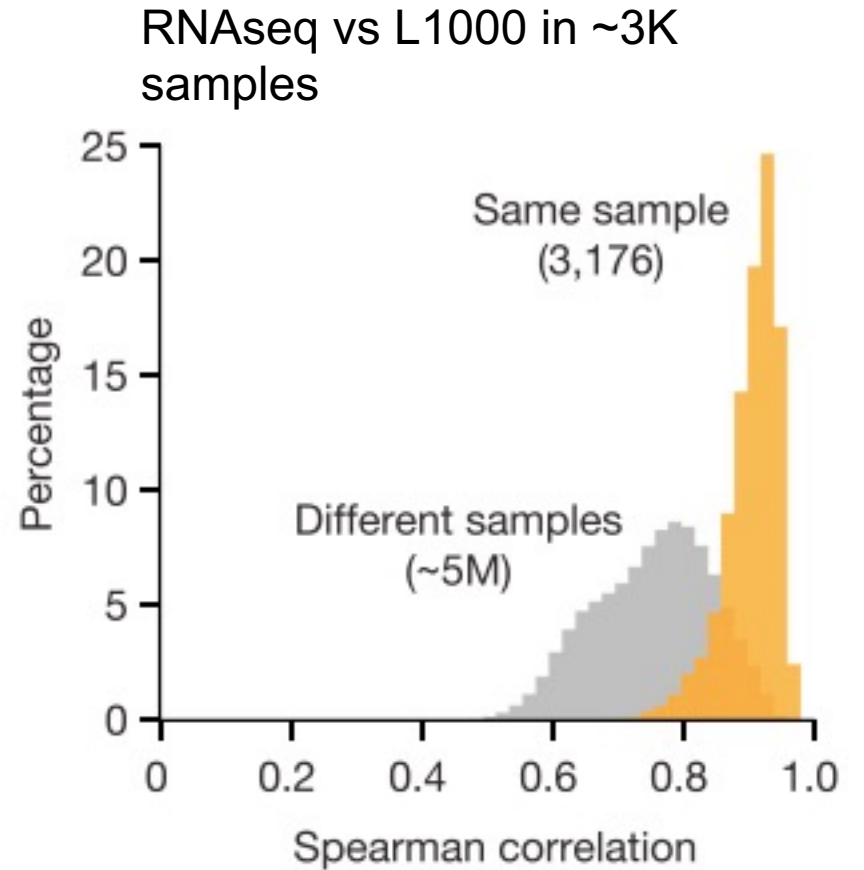
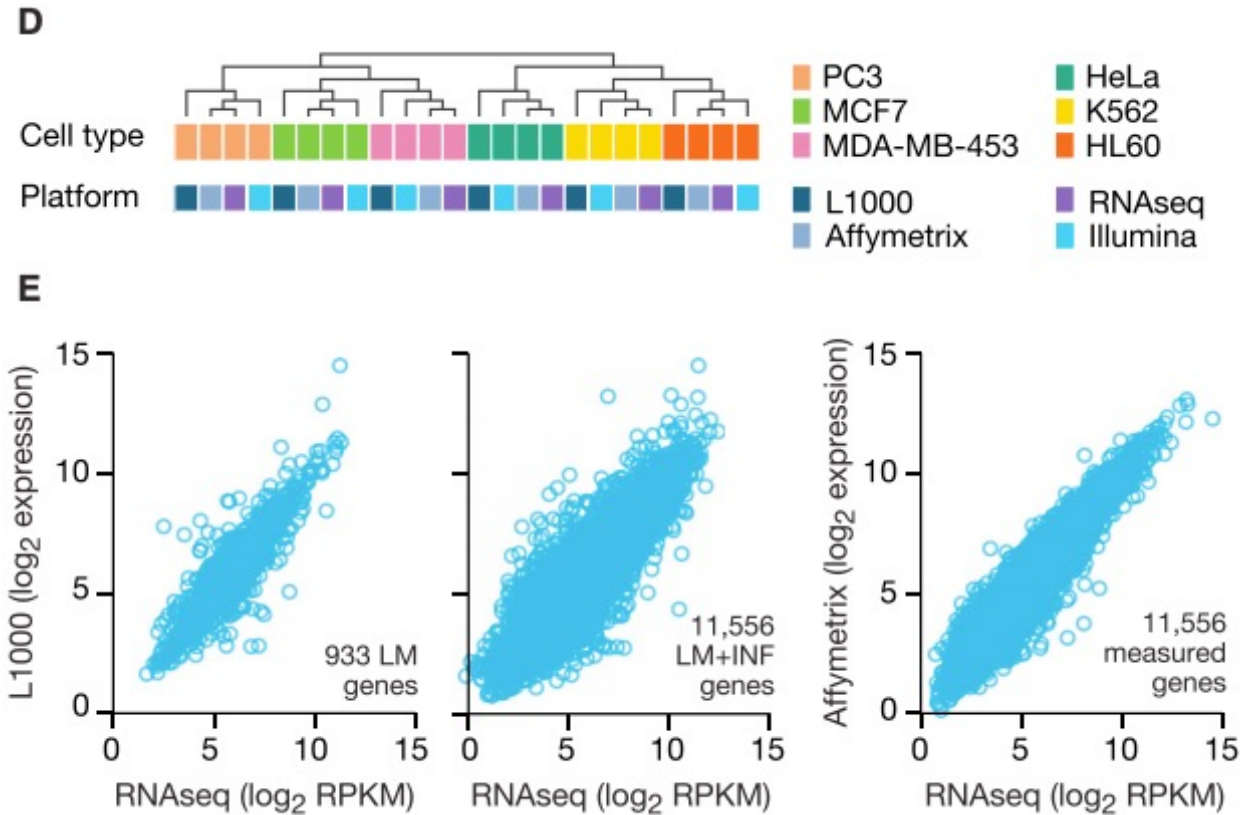
Findings from CMap pilot study

- Genomic signatures can identify drugs with common MoA
- Discover unknown MoA e.g. HDAC activity of valproic acid (initially developed as an antiseizure drug)
- Identify potential new therapeutics using a disease-associated gene query signatures
- Signatures are often conserved across diverse cell types and settings
 - Drug target needs to be expressed in that cell line e.g estrogen receptor
- Not highly sensitive to the precise concentration of drug

2nd Generation CMAP - LINCS1000

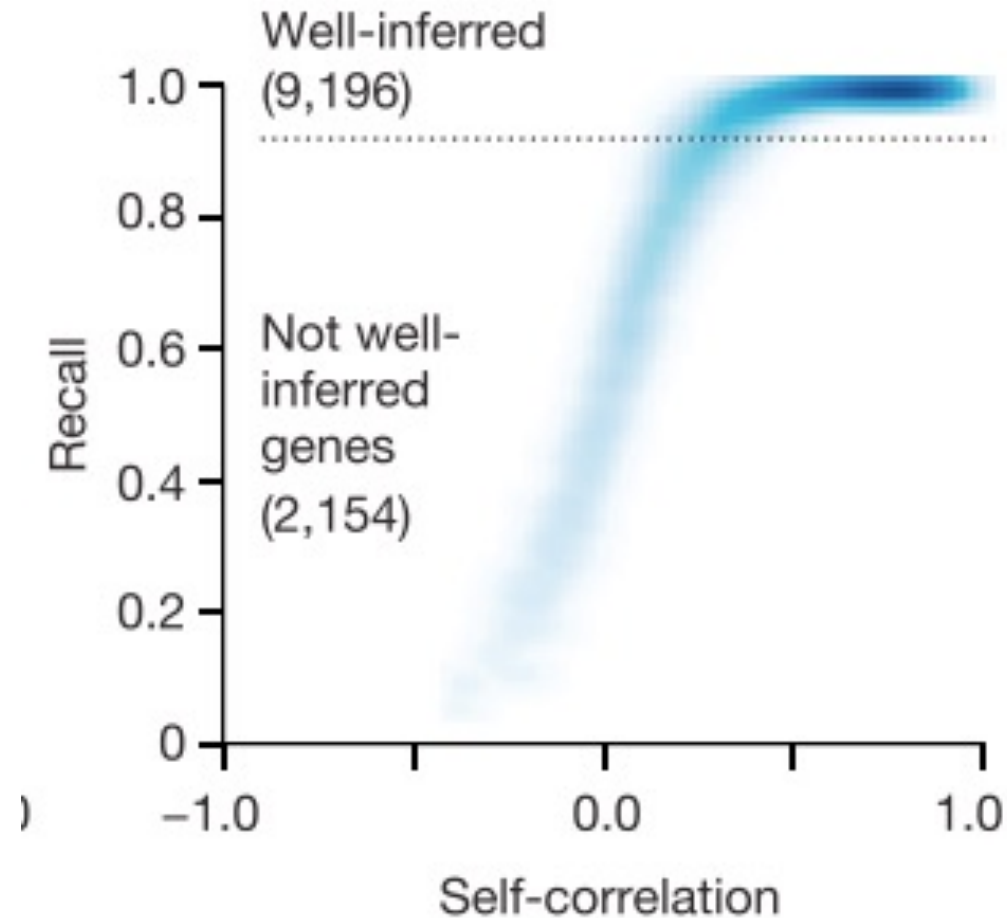
- **Library of Integrated Network-Based Cellular Signatures**
- 1000-fold scale up of the CMAP – more compounds and cell lines plus genetic perturbations.
- Capture cellular state at low cost by measuring a reduced representation of the transcriptome.
 - Analysed 12K Affy HGU133A expression profiles in GEO
 - Identified the optimal N of informative transcripts (“landmark” transcripts)
 - Cost vs information captured
 - 1000 landmarks enough to capture 82% of full transcriptome
 - No substantial enrichment of particular protein class or developmental lineage in landmark list.

Comparison of L1000 with RNAseq



strong degree of similarity of profiles across L1000 and RNA-seq platforms.

Imputation of GTEx data



~1000 landmark genes
~9200 well-inferred genes
~2000 (not well-)inferred genes

Only landmark and well-inferred genes used in analyses.

CMap-L1000v1

- 19,811 compounds profiled in triplicate (at 6 and/or 24 hrs)
- Genetic perturbation (KD or overexpression) of 5075 genes measured after 96 hrs (triplicates)
- 77 cell lines
- 470K gene signatures from ~42K perturbagens – 1000-fold increase of CMap pilot dataset.
- All data (at multiple processing levels) available in GEO (GSE92742)
- Web-based tool to query database <https://clue.io>

Generating disease gene expression signatures for querying CMap

1. Your own experiments

- Gene expression differences in cases vs controls

2. Gene Expression Omnibus



- <https://www.ncbi.nlm.nih.gov/geo/>
- Public repository of microarray, next-generation sequencing, and other forms of high-throughput functional genomic data
- Allows differential gene analysis of data
 - Select significance threshold, fold change threshold, multiple correction method
- Provides R-script for analysis

3a. Gene expression signature prediction from individual-level GWAS data using PrediXcan

- A gene-level association approach that tests the mediating effects of gene expression levels on phenotypes.
- Requires 3 datasets
 - a) GWAS data for phenotype of interest
 - b) Expression QTL training set e.g. GTEx
 - c) Population reference (e.g. 1000 Genomes)

	Trait	g1	g2	g3
ind1				
ind2				
ind3				

3a. Gene expression signature prediction from individual-level GWAS data using PrediXcan

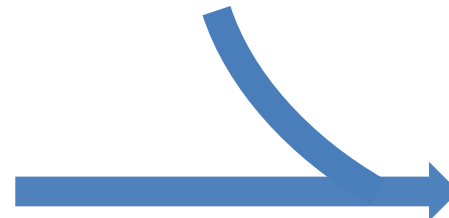
dataset1	Trait	g1	g2	g3
ind1				
ind2				
ind3				



	b	se	pval
g1			
g2			
g3			

Gene expression associated with trait

dataset 2
eQTL data,
training data for
prediction model



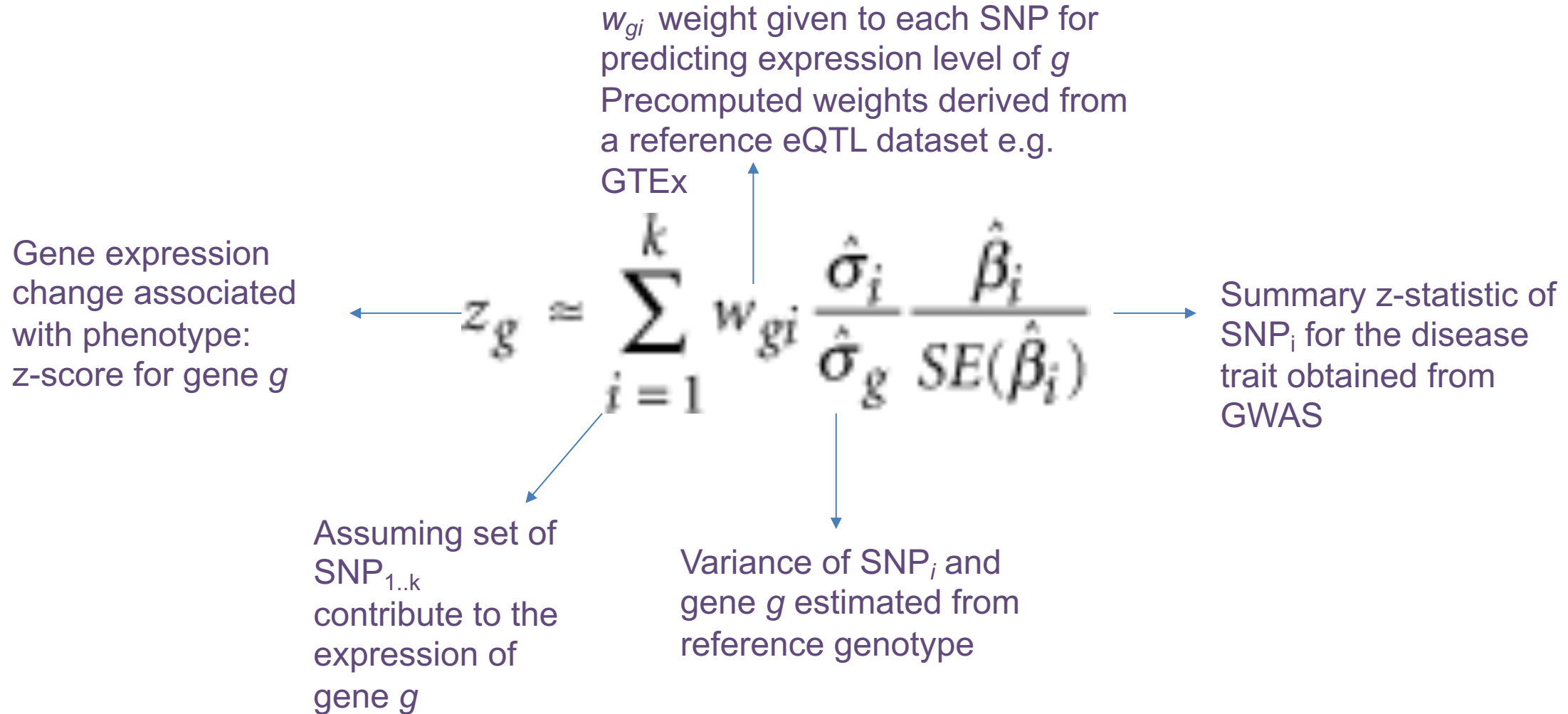
dataset1	Trait	SNP1	SNP2	SNP3
ind1				
ind2				
ind3				

	Trait	$\hat{g}1$	$\hat{g}2$	$\hat{g}3$
ind1				
ind2				
ind3				

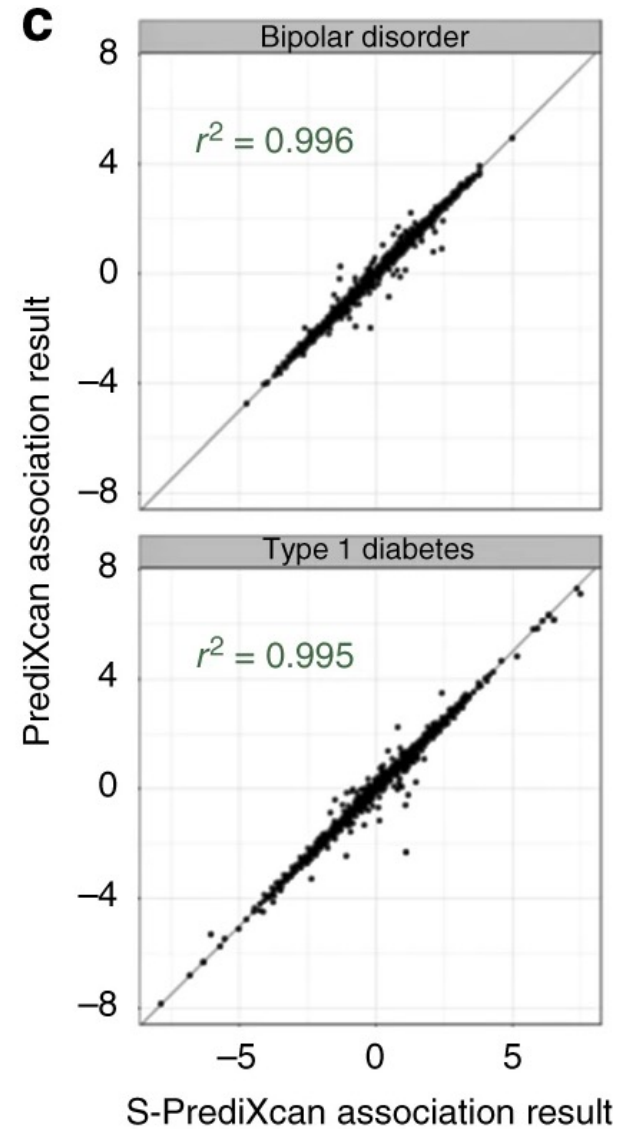
Genetically-predicted gene expression



3b. Gene expression signature prediction from GWAS summary data using S-PrediXcan



Comparison of PrediXcan and S-PrediXcan gene z-scores



Querying CMap data with iLINC
<http://www.ilincs.org/ilincs/>

Signatures i

[Search](#) [Submit a Signature](#) [Maps](#)

Submit a Signature for Connectivity Analysis

Using provided forms submit a signature in a form of a file or gene lists.

[Upload a signature](#) [Submit up and down-regulated genes](#) [Submit gene list](#)

Upload signature file and compare it with signatures library

[Select file](#) Plain text, tab delimited files only (**Sample1**), (**Sample2**), (**Sample3**), (**Sample4**), (**Sample5**).

OR

Paste a signature [example](#)

DDR1	0.656282	0.00090283
RFC2	-0.0307033	0.81855521
HSPA6	-0.0807417	0.550775065
PAX8	-2.557	2.20778E-005
GUCA1A	-0.0720556	0.545070543

[Submit signature](#)

Signature Analysis Tools 

Signature Data 

Connected Signatures  

Connected Perturbations  

Pathway Analysis

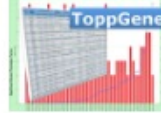
Enrichr



DAVID



ToppFun



Reactome



Background gene list very important when doing functional/pathway enrichment analysis.

Network Analysis

SPIA Analysis



GeneMANIA



X2K



SigNetA



For CMap data, background list is not all genes in the human genome, rather all genes profiles in CMap (~12,000 genes))

Visualization

PiNET



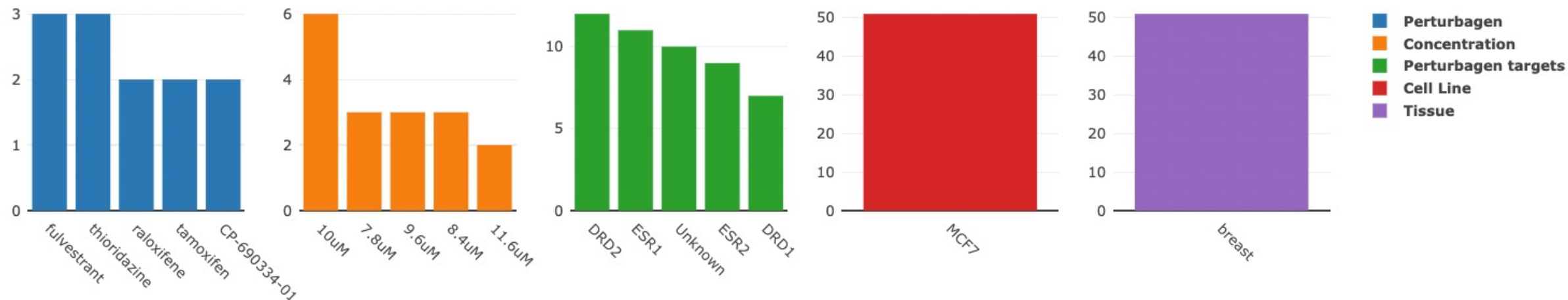
L1000FWD



▼ 51 of Connectivity Map signatures



/ Selection



Signature Id	Perturbagen	Perturbagen targets	Concentration	Cell Line	Tissue	Concordance	pValue	nGenes
<input type="checkbox"/> CMAP_127	raloxifene	ESR1 ESR2	7.8uM	MCF7	breast	1.000	0	100
<input type="checkbox"/> CMAP_128	raloxifene	ESR1 ESR2	0.1uM,7.8uM	MCF7	breast	0.943	1.5e-48	100
<input type="checkbox"/> CMAP_88	tamoxifen	ESR1 ESR2	7uM	MCF7	breast	0.922	4.5e-42	100
<input type="checkbox"/> CMAP_864	corticosterone	HSD11B1 NR3C2	11.6uM	MCF7	breast	0.917	6.2e-41	100
<input type="checkbox"/> CMAP_742	clomifene	ESR1	6.6uM	MCF7	breast	0.904	5.1e-38	100

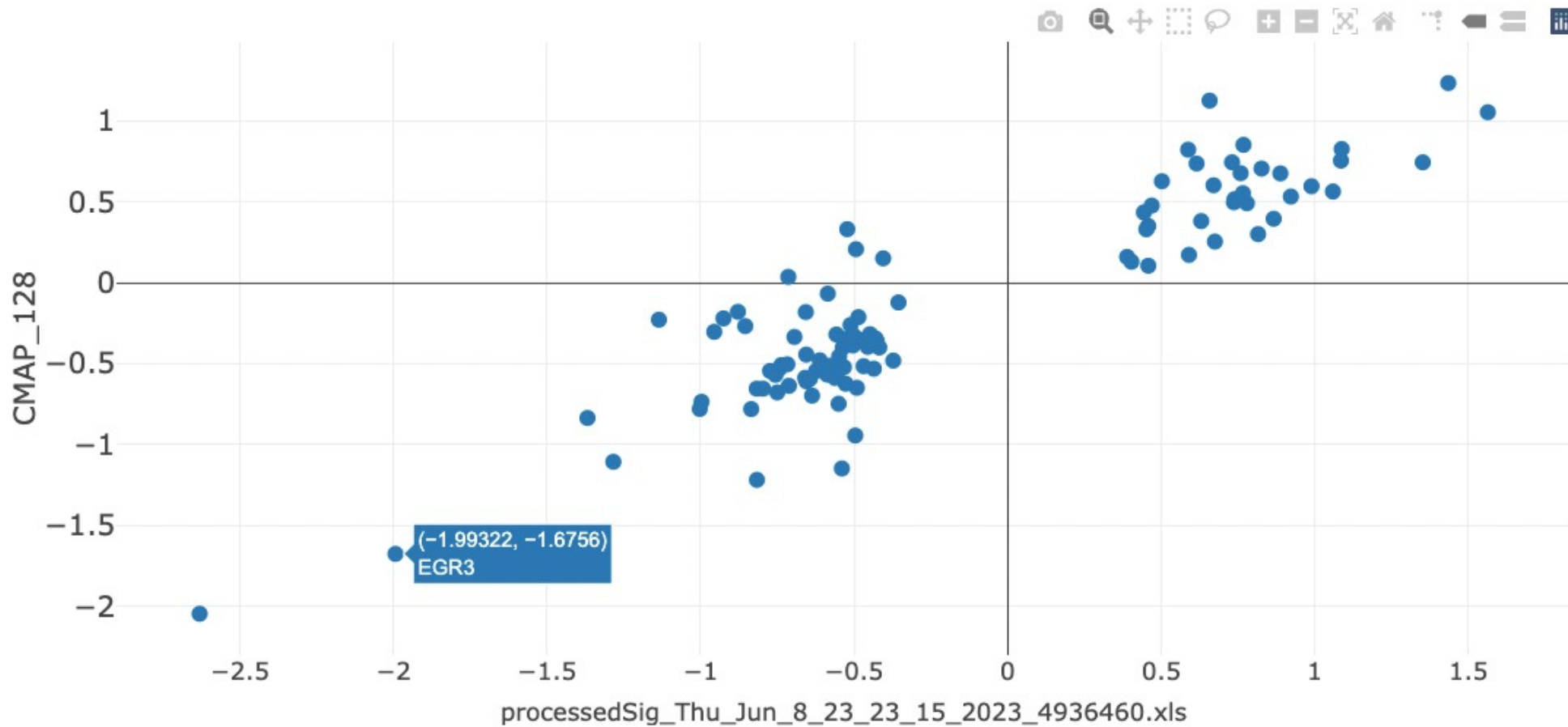
5 25 50 100

First « 1 2 3 4 5 » Last

Correlation plot

Weighted Pearson correlation: **0.943**

Pearson correlation: **0.913**



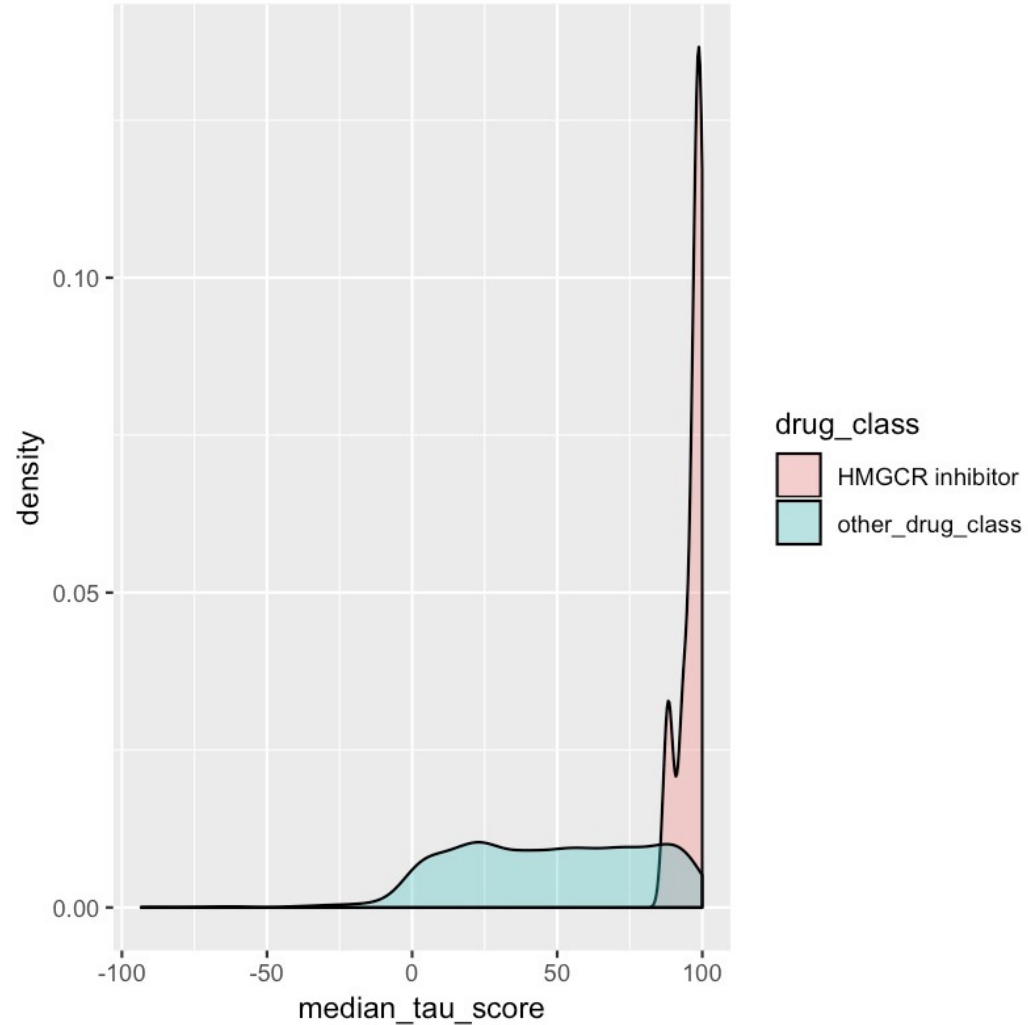
Use selected genes

Cancel

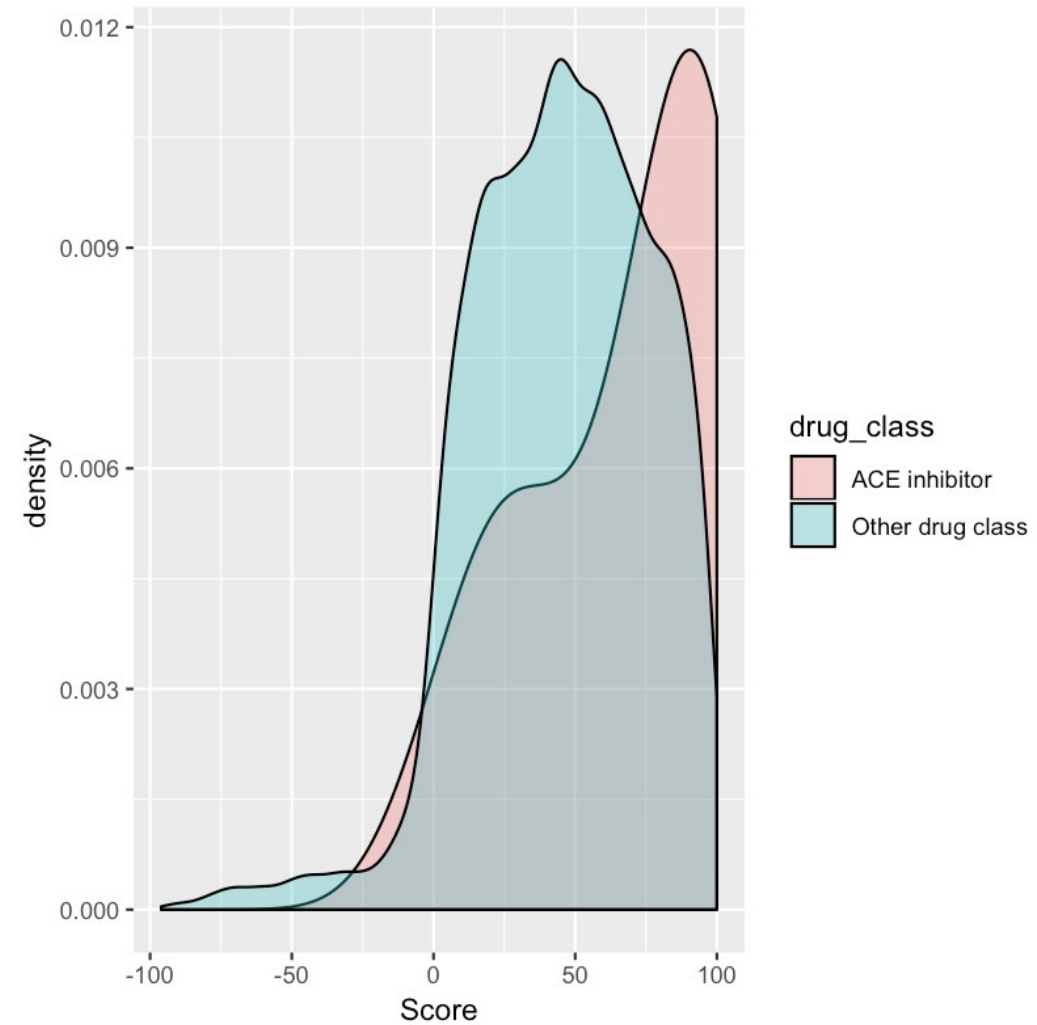
Take home messages

- iLINCS is a useful resource but requires careful manual curation
 - Check connectivity between gene knockdown/overexpression and drug
 - Check specificity of the gene signature
 - Check connectivity between compounds with same MoA

Connectivity of rosuvastatin with other HMGCR-inhibitors and all other compounds



Connectivity of enalapril with other ACE inhibitors and all other compounds



Take home messages

- iLINCS is a useful resource but requires careful manual curation
 - Check connectivity between gene knockdown/overexpression and drug
 - Check specificity of the gene signature
 - Check connectivity between compounds with same MoA
 - Check connectivity across cell lines
 - Drugs may not be in an active form. Need to check this from other sources e.g. DrugBank
 - Check if target is expressed in cell line before interpreting results (human protein atlas)

