

# UQ Genetics and Genomics Winter School 2023

Systems Genomics and Pharmacogenomics Module 6 Day 2



Gene expression signature matching for identifying drug candidates







Genetic variants

Disease genes

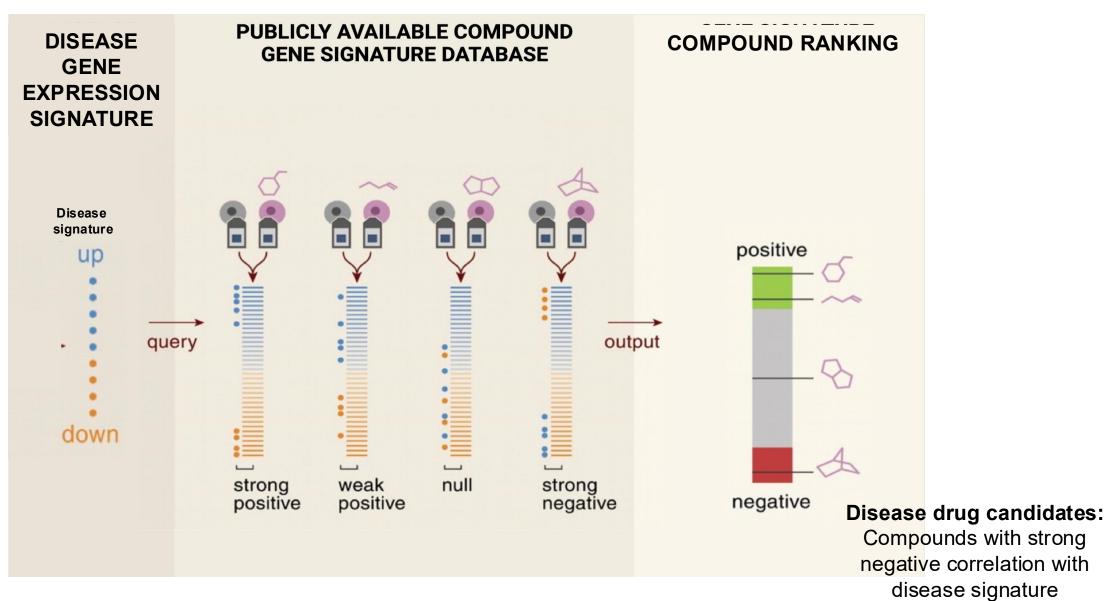
Drug candidates

- Are GWAS-significant genes targets of existing drugs (identify drug repurposing candidates)
  - Repurposing FDA-approved compounds better safety profile, lower risk, shortest path to approval
  - Can use MR approaches to prioritise genes targeted by existing drugs
- But...
  - Important disease biology may be lost under stringent p-value thresholds
  - Only considers a single gene target rather than a biological pathway
  - MR cannot be used for compounds with unknown mechanism of action (MoA)



# Gene expression signatures matching for drug discovery







# Gene expression signatures matching for understanding drug pharmacodynamics i.e. MoA

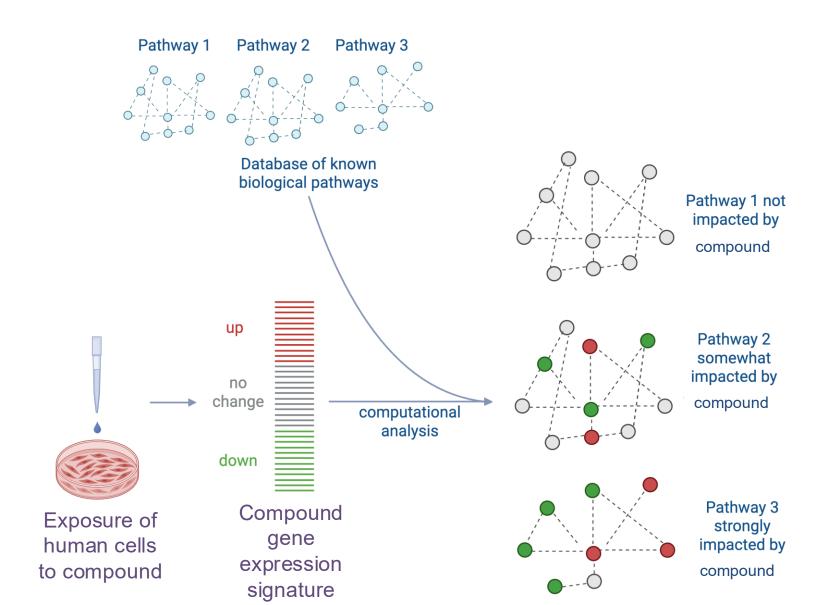
# Gene expression signature matching to understand drug pharmacodynamics



#### Approach 1: Network analysis

Which biological pathways are perturbed by your compound in human cells?

- Map these genes to their biological networks/pathways to understand which pathways are strongly impacted by the compound
- Identify "hub" genes which play a crucial role in these biological processes

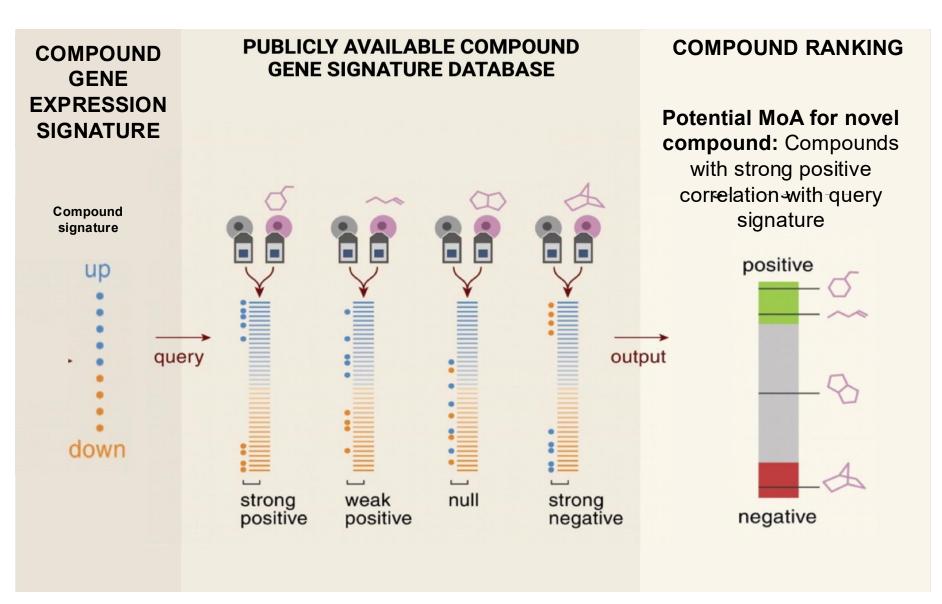


# Gene expression signature matching to understand drug pharmacodynamics



Approach 2: Comparative analysis

Which compounds with known MoA have similar signatures to your compound?



# Gene expression signature matching for drug discovery



- 1) A database of gene expression signatures for drugs
- 2) A disease gene expression signature
- 3) Query the signature database using the disease gene expression signature to identify compounds that <u>'reverse</u>' disease gene expression changes.

Does not require knowledge of the drug's MoA

Does not require an understanding of disease pathophysiology

# Gene expression signature matching for drug pharmacodynamics



- 1) A database of gene expression signatures for drugs
- 2) Novel compound gene expression signature easily done using compound perturbation studies using cells.
- 3) Use network or comparative analysis (latter requires database of compound signatures)



## Connectivity Map (CMap)

Library of gene expression signatures in response to chemical and genetic perturbation.

- >1 million gene expression profiles
- ~50 different cell lines
- ~20,000 compounds (chemical perturbation)
- ~5,000 knockdown/overexpression (genetic perturbations)



RESEARCH ARTICLES

#### The Connectivity Map: Using Gene-Expression Signatures to Connect Small Molecules, Genes, and Disease

JUSTIN LAMB, EMILY D. CRAWFORD, DAVID PECK, JOSHUA W. MODELL, IRENE C. BLAT, MATTHEW J. WROBEL, JIM LERNER, JEAN-PHILIPPE BRUNET, ARAVIND SUBRAMANIAN

https://www.broadinstitute.org/connectivity-map-cmap

### THE UNIVERSITY OF QUEENSLAND AUSTRALIA

# 1st Generation CMap - Lamb et al Science 2006

- Need to establish the relation among diseases, physiological processes, and the action of small-molecule therapeutics.
- Previous compound and genetic perturbation studies in yeast and rats
  - Translation to humans
  - High cost of animal studies
- Mammalian cells
  - Generalisable, systematic and biologically relevant
  - BUT...a large number of parameters would need to be optimized for each perturbation cell type, dose, duration
- Pilot study demonstrated the feasibility of this approach



## 1<sup>st</sup> Generation CMap - compounds

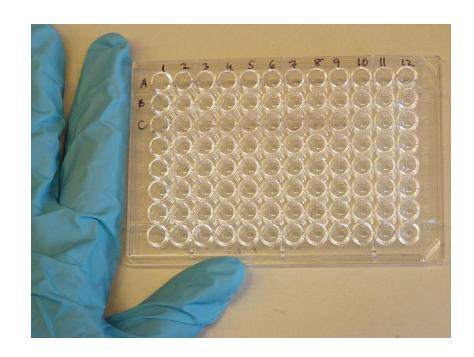
164 distinct small-molecule perturbagens, selected to represent a broad range of activities:

- FDA—approved drugs
- nondrug bioactive "tool" compounds
- multiple compounds sharing molecular targets (test if they share gene signatures e.g. HDAC inhibitors)
- compounds with the same clinical indication (test whether compounds with different MoA that treat the same disease generate similar gene signatures e.g. antidiabetics)
- Molecules that are proximal (e.g. selective estrogen receptor modulators) and distal to gene expression
- Molecules whose targets are not expressed in the cell types being tested (COX2 inhibitors)





- Stably grown over long periods of time
- Amenable to culture in microtiter plates
- breast cancer epithelial cell line MCF7
  - extensively molecularly characterised,
  - used as a reference cell line
- prostate cancer epithelial cell line PC3
- nonepithelial lines HL60 (leukemia) and SKMEL5 (melanoma)
- Assess degree to which gene signatures are context-dependent





### 1st Generation CMAP – dose and duration

- 10uM optimal concentration is not known for many compounds
  - Toxicity studies required for proper optimisation of dose
- 6 and 12 hrs post-treatment
  - Profiles obtained too early might not yield robust signals—esp for perturbations that do not directly modulate transcription
  - Profiles obtained too late may reflect secondary and tertiary responses
  - obtain signatures related to direct mechanisms of action
- Dose and duration dependent on question of interest, but difficult to optimise in such high-throughput experiments.



## Compound gene signature generation

- Control perturbations for each treatment (cells grown on the same plate treated with vehicle only)
  - minimize the impact of batch-to-batch
  - biological and technical variation
- Replicates
- Data were collected in multiple batches over a period of 1 year by Affymetrix GeneChip microarrays.
- DEG analysis compound-treated gene expression vs intra-batch vehicle-treated control
- For each treatment ~22,000 genes rank-ordered according to differential expression

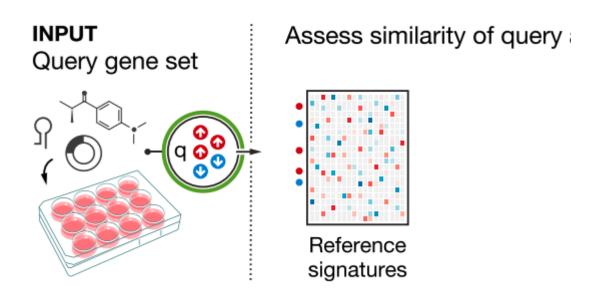


Can gene expression signature matching

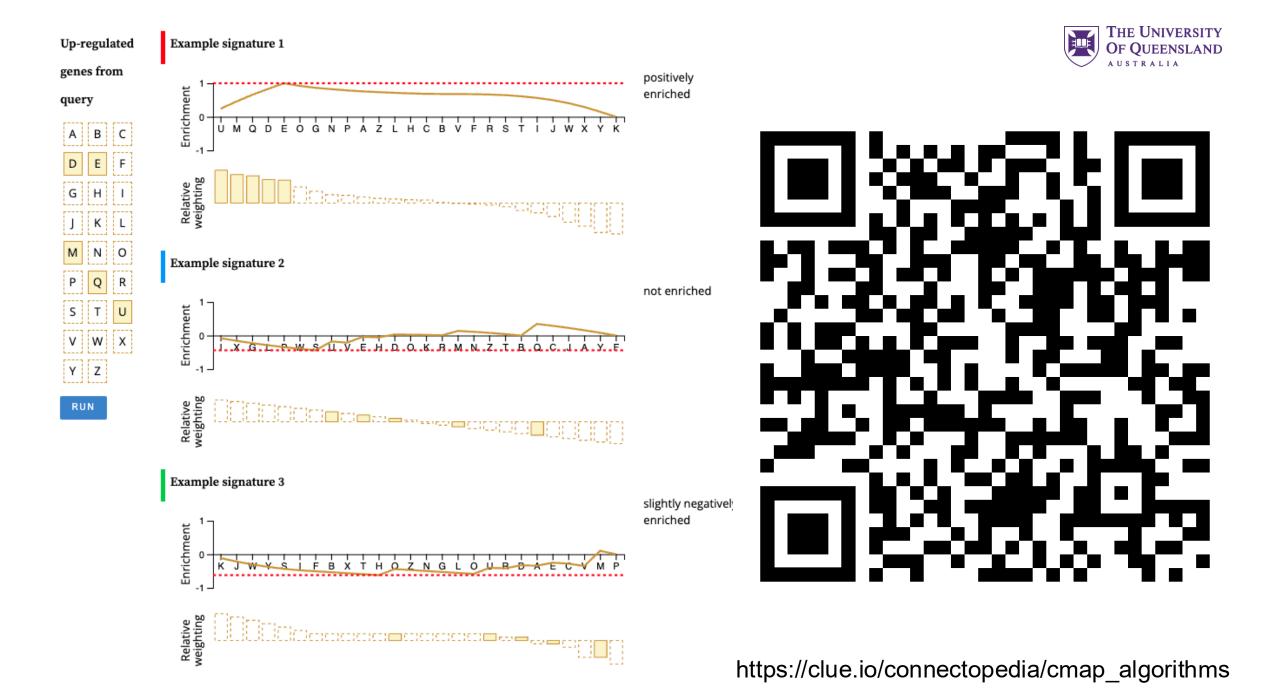
- a) identify MoA of a compound?
- b) identify drug candidates for disease?



## Connectivity score - metric for signature similarity

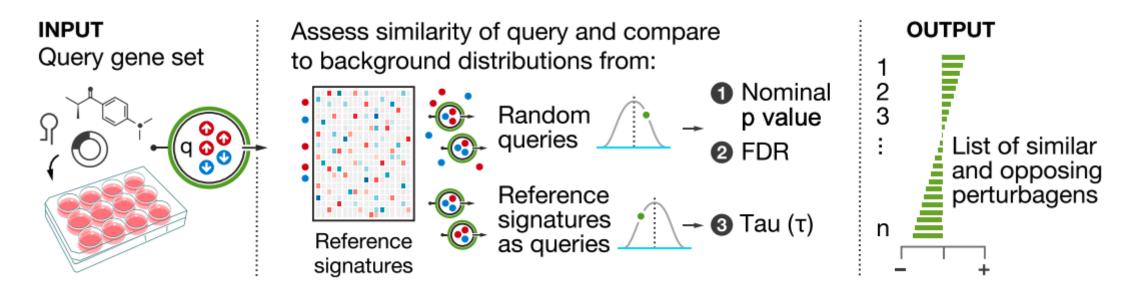


- Rank-based pattern-matching strategy based on the Kolmogorov-Smirnov statistic
- Determine if the most significant DEGs in query gene set are randomly distributed in the reference compound signature
- Enrichment score reflects the degree to which your query gene set is overrepresented in the extremes of the ranked reference gene signature





## Connectivity score - metric for signature similarity



#### Measures of confidence:

- Nominal p-value comparing similarity of query and reference signature to null distribution of random queries, using KS enrichment statistic
- Tau score compares an observed enrichment score to all others in the database a standardized measure ranging from -100 to 100. A Tau of 90 indicates that only 10% of signatures in the database had a stronger connectivity to the query than the compound in question.

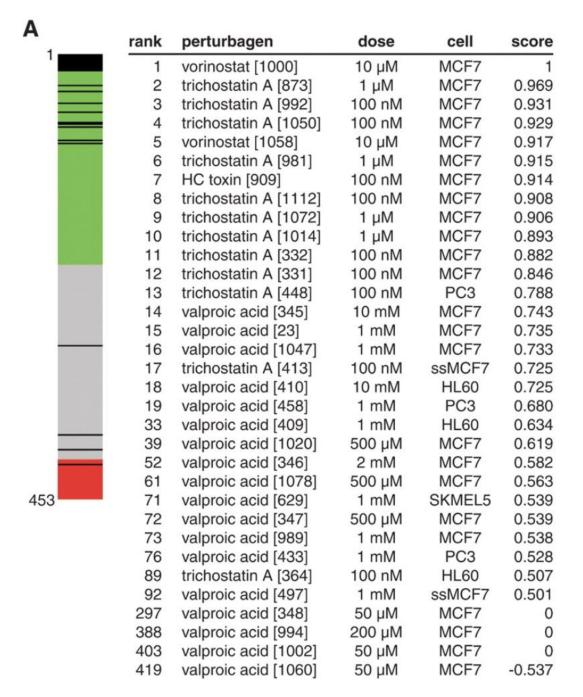
https://clue.io/connectopedia/cmap\_algorithms



# Example results – HDAC inhibitors

- HDACs remove acetyl groups on histones and regulate gene expression
- Query HDAC signature derived from independent study:
  - response of bladder and breast cancer cells treated with 3 HDAC inhibitors (vorinostat, MS-27-275, trichostatin)
  - 13-gene (8 up and 5 down-regulated) signature

• Determine if a query signature can recover compounds from the same class (same MoA).



- Compounds with HDAC inhibitory effects shown by black lines
- Despite differences in cell lines used to generate query signature, the approach identifies HDAC inhibitors as the top scoring compounds.
- Not highly sensitive to concentrations
- Strong connectivity with two structurally distinct compounds, valproic acid (developed as an antiseizure drug) and HC toxin, both now known to have HDAC-inhibitory activity

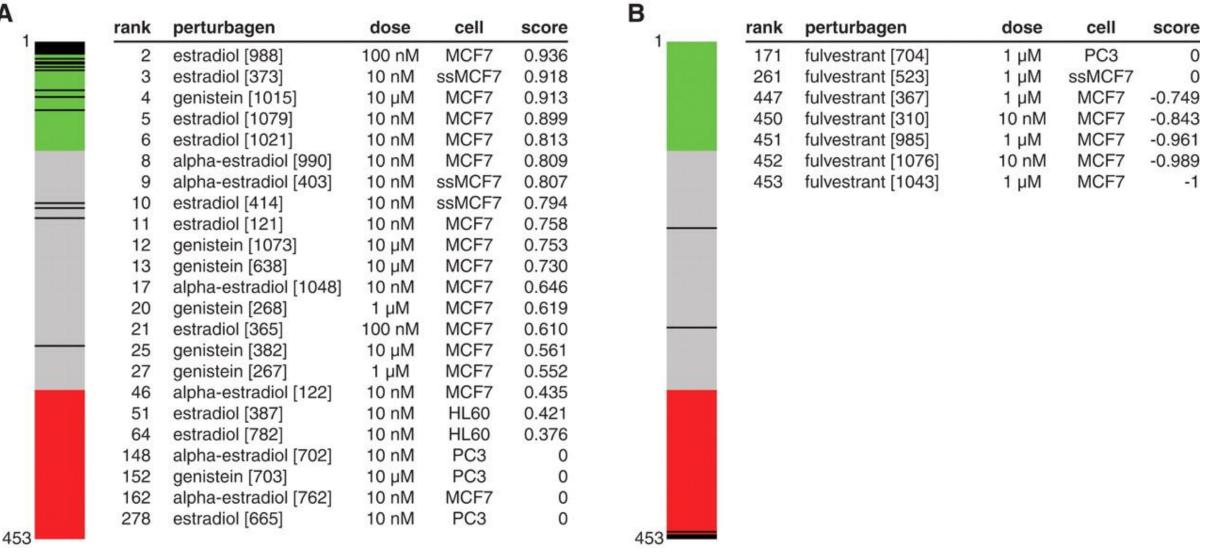
(green, positive; gray, null; red, negative



# Example - Estrogens

- Estrogen modulates nuclear hormone signaling by binding to estrogen receptor.
- Query signature from an independent experiment MCF7 cells treated with 17beta-estradiol
  - 129-gene signature (40 up and 89 down-regulated)





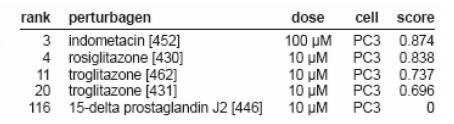
no robust connections recovered in PC3 or HL60 cells, neither of which expresses ER.

highest negative connectivity scores came from fulvestrant, a known anti-estrogenic drug



## Connections with Disease States

- Query DEGs from a rat model of diet-induced obesity
- Several differences in exp design:
  - · Species: Rat vs human,
  - Exposure duration: 65 days vs 6 hrs
  - Tissue: adipose vs cancer cell line
- 3 PPAR-gamma agonists identified
- PPAR-gamma agonists are known potent inducers of adipogenesis in vitro
- Troglitazone and rosiglitazone are anti-diabetic treatments, with weight-gain as a known major side effect
- BUT...null or negative scores in non-PC3 cell lines, (only PC3 expresses PPAR-gamma)



# Findings from CMap pilot study



Gene expression signatures can

- 1. Identify drugs with common MoA
- 2. Identify unknown MoA of drugs
- 3. Identify potential new therapeutics for disease
- 4. Are often conserved across diverse cell types and settings
  - Drug target needs to be expressed in that cell line
- 5. Not highly sensitive to the precise concentration of drug



# 2<sup>nd</sup> Generation CMAP - LINCS1000

- Library of Integrated Network-Based Cellular Signatures
- 1000-fold scale up of the CMAP more compounds and cell lines plus genetic perturbations.

- Gene arrays and RNAseq not suitable for large-scale profiling
  - High cost
  - RNAseq cannot detect low abundant transcripts without deep sequencing which is costly

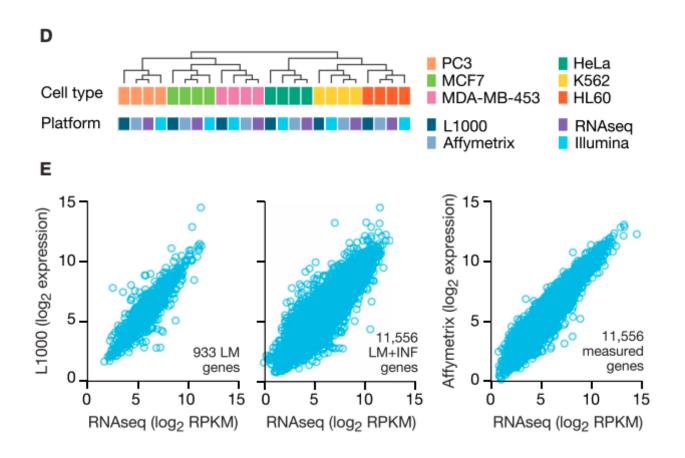


# 2<sup>nd</sup> Generation CMAP - LINCS1000

- Capture cellular state at low cost by measuring a reduced representation of the transcriptome.
- Analysed 12K Affy HGU133A expression profiles in GEO
  - Identified the optimal number of informative transcripts ("landmark" transcripts)
  - Cost vs information captured
  - 1000 landmarks enough to capture 82% of full transcriptome
- Tested ability of different number of landmark genes to recover connections observed in pilot data (for 25 signatures)
- No substantial enrichment of particular protein class or developmental lineage in landmark list (some generic classes enriched e.g. enzyme binding, ATP binding).



# Comparison of L1000 with RNAseq



strong degree of similarity of profiles across L1000 and RNA-seq platforms



# **Using CMap Data**



### CMAP – One dataset several names

### CMap-L1000 version 1:

- L1000-based compendium
- Phase 1
- A Next Generation Connectivity Map: L1000 platform and the first 1,000,000 profiles (2017)
- Available at GSE92742

#### Data:

### 19,811 compounds (drugs and small molecules):

- Different time (6 hours or 24 hours)
- Different concentration (0.04uM to 90uM)
- Different cell lines: (71 different cell lines)

### 1,319,138 replicates measured.

This dataset also include genetic perturbation



### CMAP – One dataset several names

#### CMap-L1000 version 2:

- Phase 2
- No paper published on this dataset (but the data was included in clue.io).
- Available at GSE70138

#### Data:

#### 1,768 additional compounds:

- Different time (3hours, 6 hours or 24 hours)
- Different concentration (0.04uM to 40uM)
- Different cell lines: (30 different cell lines)

### 354,123 replicates measured



### CMAP – One dataset several names

#### iLincs:

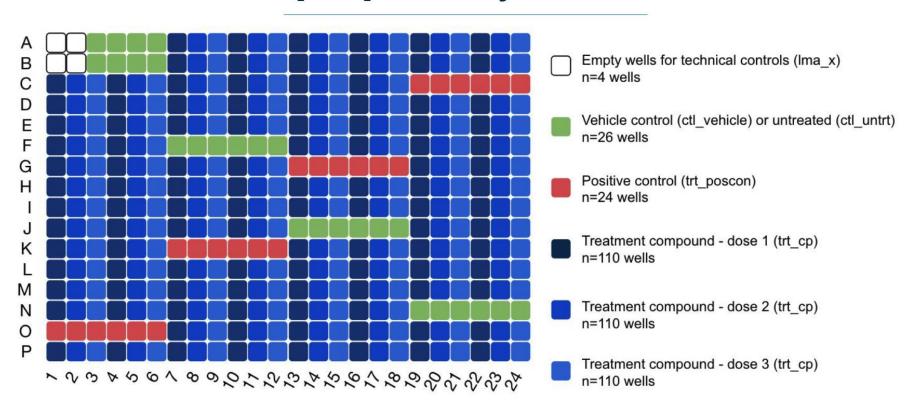
- Uses both GSE92742 and GSE70138.
- This is a meta-analysis combining phase 1 and phase 2.
- Paper: Connecting omics signatures and revealing biological mechanisms with iLINCS (2022)
- Data is accessed through the ilincs website

### 1,673,261 replicates measured

# CMAP – What does it actually look like?

### **Example plate layout: 3 dose**

**OF QUEENSLAND** 





# CMAP – Microarray.

### **Definition of the landmark genes:**

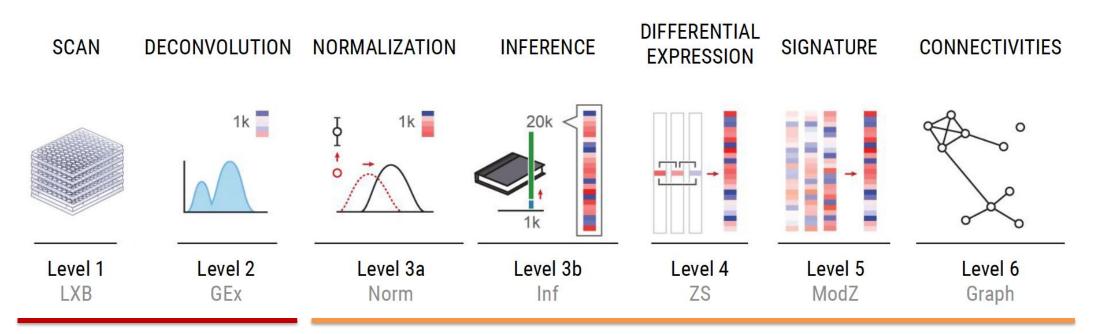
• 12,063 gene expression samples profiles using Affymetrix HG-U133A microarrays from the Gene Expression Omnibus (GEO) called  $DS_{GEO}$ .



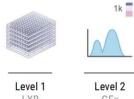


## **L1000 Data Processing**

**Processing Stages Post-Detection** 



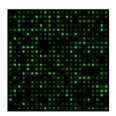
# Microarray processing:

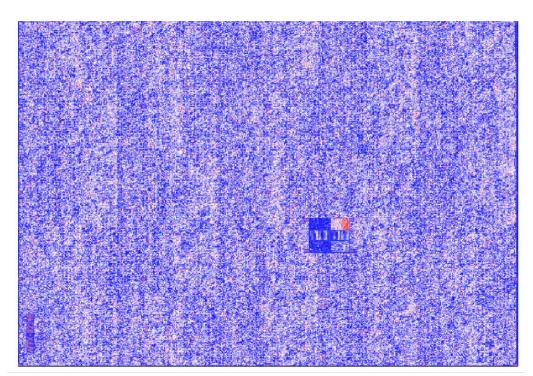


SCAN









Example Slide Affymetrix microarray (~18,000 genes)

Data generation for each of the replicates:

- Each dot on the slide corresponds to a different gene.
- The intensity and color of the dot represent:
  - Blue indicates low signal intensity (low expression or hybridization).
  - Red indicates high signal intensity (high expression or hybridization).
  - Intermediate shades (purple, magenta) indicate medium signal.

The centre of the image corresponds to control genes of known intensity. This allows to generate a numerical value for each gene measured.



# CMAP processing - Level 3 normalisation

NORMALIZATION

Level 3a

Norm

At this stage, the data contain only 1,058 genes measured for each replicate (1,673,261)

Normalisation is called L1000 Invariant Set Scaling.

For each sample the expression of 80 invariable gene is used to generate a "calibration curve"



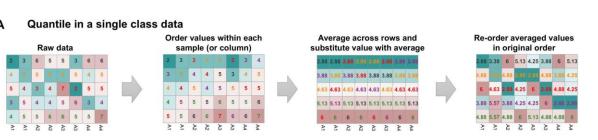
The data is then recalibrated using the following equation:

$$y_{scaled} = ay_{raw}^b + c$$

### Where:

- a,b and c are estimated within each sample using a least square approach.
- $y_{raw}$  is the unscaled data.
- $y_{scaled}$  is the scaled data used for further analysis.

 $y_{scaled}$  is the normalized using a quantile normalization.

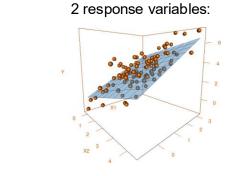


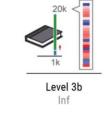
# CMAP processing - Level 3 gene inference

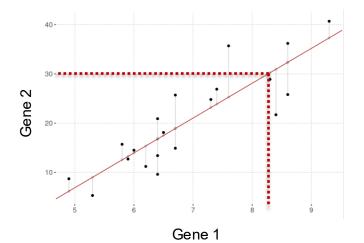
INFERENCE

To infer the value of the missing genes, they assume that unmeasured genes can be predicted from the measured landmark genes using the following linear combination:

$$x = w_0 + \sum_{i=1}^{978} w_i y_i$$







Example:

Gene 1: Measured value of 8.2 Gene 2: Predicted value of 30

| Gene Symbol | Gene Title   | Self-Correlation | Feature set |
|-------------|--|------------------|-------------|
| ESRRA       | estrogen related receptor alpha                      | 0.89             | BING        |
| EIF3D       | eukaryotic translation initiation factor 3 subunit D | 0.90             | BING        |
| HAUS2       | HAUS augmin like complex subunit 2                   | -0.38            | Inferred    |

# CMAP processing – Level4: Z-score scaling

DIFFERENTIAL EXPRESSION

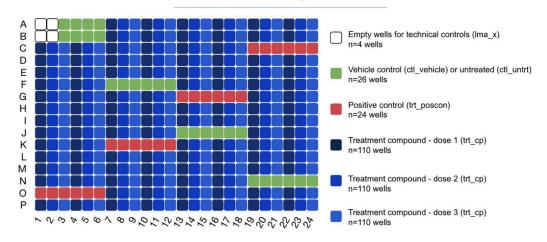
To make genes comparable, they are changed to a z-score scale using the following formula:

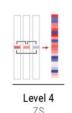
$$z_i = \frac{y_{norm} - \mu}{\sigma}$$

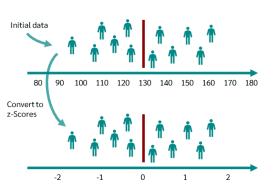
### Where:

- $z_i$  is the z-score transformed expression value.
- y<sub>norm</sub> is the normalized expression (measured or inferred)
- $\mu$  is the mean normalized expression on the plate
- $\sigma$  is the standard deviation of the normalized expression on the plate

### **Example plate layout: 3 dose**









## CMAP processing – Level 5: consensus signatures

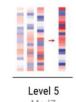
- SIGNATURE
- Level 5
- Level 5 ModZ

- Pairwise correlation is calculated between each replicates of a signature
  - Drug
  - Time of expose
  - Dose
  - Cell line
- The consensus signature is then calculated as the linear combination of the replicates gene expression.
  - The coefficients are the sum of its correlation to the other replicates normalized to sum to 1.

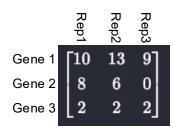


## CMAP processing – Level 5: consensus signatures

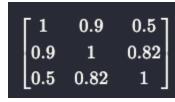
SIGNATURE



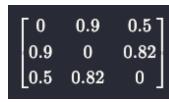
### Example 3 genes, 3 replicates:



### Step 1: correlation matrix:



### Step 2: Set self-correlation to 0:



### Step 3: Raw weights:

### Step 4: Normalized weights:

### Step 5: Linear combination

$$\begin{bmatrix} 0.31*10+0.39*13+0.30*9\\ 0.31*8+0.39*6+0.30*0\\ 0.31*2+0.39*2+0.30*2 \end{bmatrix} = \begin{bmatrix} 10.87\\ 4.82\\ 2 \end{bmatrix}$$
 Consensus Signature



# Generating disease gene expression signatures for querying CMap

## 1. Gene Expression Omnibus





- https://www.ncbi.nlm.nih.gov/geo/
- Public repository of microarray, nextgeneration sequencing, and other forms of high-throughput functional genomic data
- Allows differential gene analysis of data
  - Select significance threshold, fold change threshold, multiple correction method
- Provides R-script for analysis

| Browse Content     |         |  |  |
|--------------------|---------|--|--|
| Repository Browser |         |  |  |
| DataSets:          | 4348    |  |  |
| Series:            | 202182  |  |  |
| Platforms:         | 25116   |  |  |
| Samples:           | 5887793 |  |  |

#### **Series GSE11393**

### Query DataSets for GSE11393

Status Public on Jun 03, 2008

Title Monocyte gene expression profiling in familial combined hyperlipidemia and its

modification by atorvastatin treatment

Organism Homo sapiens

Experiment type 

Expression profiling by array

Summary Introdu

Introduction: The genetic origin of familial combined hyperlipidemia (FCH) is not well understood. We used microarray profiling of peripheral blood monocytes to search novel genes and pathways involved in FCH. Methods: Fasting plasma for determination of lipid profiles, inflammatory molecules, and adipokines was obtained and peripheral blood monocytes were isolated from male FCH patients basally and after 4 weeks of atorvastatin treatment. Sex-, age- and adiposity-matched controls were also studied. Gene expression profile was analyzed using Affymetrix Human Genome U133A 2.0 GeneChip arrays. Results: Analysis of gene expression by cDNA microarrays showed that 82 genes were differentially expressed in FCH monocytes compared to controls. Atorvastatin treatment modified the expression of 87 genes. Changes in the expression of some genes, confirmed by real time RT-PCR, (CD36, leucine-rich repeats and immunoglobulin-like domains-1, tissue factor pathway inhibitor 2, myeloid cell nuclear differentiation antigen tumor necrosis factor receptor superfamily, member 25 and CD96) may be related to a proinflammatory environment in FCH monocytes, which is partially reversed by atorvastatin. Higher plasma levels of triglycerides and free fatty acids and lower levels of adiponectin in FCH patients could also trigger changes in gene expression that atorvastatin cannot modify. Conclusions: Our results demonstrate clear differences in gene expression in FCH monocytes compared with those of matched healthy controls, some of which are influenced by atorvastatin treatment.

Keywords: comparative study differential gene expression

Overall design

Peripheral blood monocytes were isolated from male FCH patients basally and after 4 weeks of atorvastatin treatment. Sex-, age- and adiposity-matched





Platforms (1) GPL571 [HG-U133A\_2] Affymetrix Human Genome U133A 2.0 Array

Samples (9) GSM287664 Monocyte control rep 1

GSM287665 Monocyte control rep 2

GSM287666 Monocyte control rep 3

**Relations** 

BioProject PRJNA106517

### Analyze with GEO2R

| Download family                 | Format   |
|---------------------------------|----------|
| SOFT formatted family file(s)   | SOFT ?   |
| MINiML formatted family file(s) | MINIML 🔃 |
| Series Matrix File(s)           | TXT 🕐    |

| Supplementary file | Size    | Download       | File type/resource |
|--------------------|---------|----------------|--------------------|
| GSE11393 RAW.tar   | 17.6 Mb | (http)(custom) | TAR (of CEL)       |



**GEO** accession

GSE11393 Set

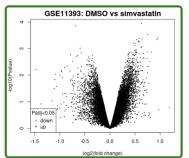
Monocyte gene expression profiling in familial combined hyperlipidemia and its modification by atorvastatin treatment

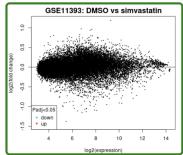
|                 |            | ▶ Define gro           | ups -  | elected 6 out of 9 s | sam   |
|-----------------|------------|------------------------|--|----------------------|-------|
|                 |            |                        | Columns  | ,                    | -   5 |
| Group Acce      | cession \$ | Title ♦                | Source name  | Characteristics      |       |
| - GSM           | M287664    | Monocyte control rep 1 | Monocyte human control sample  | Human healthy ma     | le s  |
| - GSN           | M287665    | Monocyte control rep 2 | Monocyte human control sample  | Human healthy ma     | le s  |
| - GSN           | M287666    | Monocyte control rep 3 | Monocyte human control sample  | Human healthy ma     | le s  |
| DMSO GSN        | M287667    | Monocyte FCH rep 1     | Monocyte human familial combined hyperlipidemia (FCH) sample                                   | Human male subje     | cts   |
| DMSO GSN        | M287668    | Monocyte FCH rep 2     | Monocyte human familial combined hyperlipidemia (FCH) sample                                   | Human male subje     | cts   |
| DMSO GSN        | M287669    | Monocyte FCH rep 3     | Monocyte human familial combined hyperlipidemia (FCH) sample                                   | Human male subje     | cts ' |
| simvastatin GSM | M287670    | Monocyte ATV rep 1     | Monocyte human familial combined hyperlipidemia after treatment with atorvastatin (ATV) sample | Human male subje     | cts   |
| simvastatin GSM | M287671    | Monocyte ATV rep 2     | Monocyte human familial combined hyperlipidemia after treatment with atorvastatin (ATV) sample | Human male subje     | cts   |
| simvastatin GSM | M287672    | Monocyte ATV rep 3     | Monocyte human familial combined hyperlipidemia after treatment with atorvastatin (ATV) sample | Human male subje     | cts   |

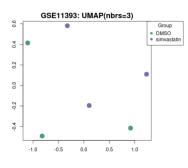


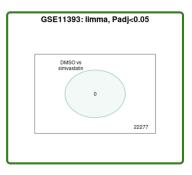
Reanalyze if you changed any options.

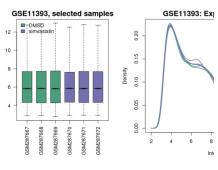
### Visualization ?











### Top differentially expressed genes ?

#### Download full table Select columns

| ID          | adj.P.Val | P.Value  | t    | В     | logFC | Gene.symbol    | Gene.title         |
|-------------|-----------|----------|------|-------|-------|----------------|--------------------|
| 214551_s_at | 0.65      | 0.000114 | 7.83 | -2.37 | 0.624 | CD7            | CD7 molecule       |
| ▶ 201042_at | 0.65      | 0.000137 | -7.6 | -2.39 | -0.69 | TGM2           | transglutaminase 2 |
| 204612_at   | 0.65      | 0.000297 | 6.71 | -2.5  | 0.754 | PKIA           | protein kinase (cA |
| ▶ 206761_at | 0.65      | 0.0004   | 6.38 | -2.54 | 0.956 | CD96           | CD96 molecule      |
| ▶ 201323_at | 0.65      | 0.000412 | 6.35 | -2.55 | 0.58  | MIR6733///EBNA | microRNA 6733///   |

GEO2R

Options

Profile graph

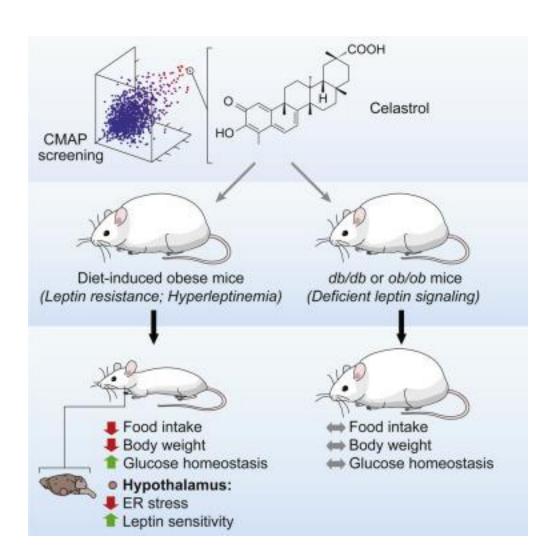
R script

```
# Version info: R 4.2.2, Biobase 2.58.0, GEOquery 2.66.0, limma 3.54.0
Differential expression analysis with limma
library(GEOquery)
library(limma)
library(umap)
# load series and platform data from GEO
gset <- getGEO("GSE11393", GSEMatrix =TRUE, AnnotGPL=TRUE)</pre>
if (length(gset) > 1) idx <- grep("GPL571", attr(gset, "names")) else idx <- 1</pre>
gset <- qset[[idx]]</pre>
# make proper column names to match toptable
fvarLabels(gset) <- make.names(fvarLabels(gset))</pre>
# group membership for all samples
gsms <- "XXX000111"
sml <- strsplit(gsms, split="")[[1]]</pre>
# filter out excluded samples (marked as "X")
sel <- which(sml != "X")</pre>
sml <- sml[sel]</pre>
gset <- gset[ ,sel]</pre>
# log2 transformation
```



# 2. Animal experiments





### Liu et al 2015 Cell

- Aim: Identify potential drug candidates for reducing ER stress and obesity
- Endoplasmic reticulum (ER) stress tightly linked to obesity.
- Gene signature Induced ER stress in mice and measured transcriptional response in the livers of these mice.
- Query CMAP Celastrol (extract from Thunder god vine plant)
- Validation in mice studies Celastrol increases leptin sensitivity to suppress food intake in mice.
- ERX Pharmaceuticals (founded in 2014) currently testing leptin sensitisers in Phase I clinical trials.



# 3. Human gene expression studies

### Gene expression differences in human cases vs controls

| dataset1 | Trait | g1 | g2 | g3 |
|----------|-------|----|----|----|
| ind1     | 1     |    |    |    |
| ind1     | 0     |    |    |    |
| ind2     | 1     |    |    |    |

Measure gene expression levels in cases (1) and controls (0)

Association between gene expression and disease status

- May be hard to get DEGs in different tissues

# 4a. From individual-level GWAS data using PrediXcan



b se pval
g1
g2
g3

Gene
expression
associated
with trait

dataset 2
eQTL data,
training data for
prediction model



|      | Trait | ĝ1 | <b>ĝ2</b> | ĝ3 |
|------|-------|----|-----------|----|
| ind1 | 1     |    |           |    |
| ind2 | 0     |    |           |    |
| ind3 | 1     |    |           |    |

Genetically-predicted gene expression

| dataset1 | Trait | SNP1 | SNP2 | SNP3 |
|----------|-------|------|------|------|
| ind1     | 1     |      |      |      |
| ind2     | 0     |      |      |      |
| ind3     | 1     |      |      |      |



### 4b. From GWAS summary data using S-PrediXcan

- Requires 3 datasets
  - a) GWAS data for phenotype of interest
  - b) Expression QTL training set e.g. GTEx
  - c) Population reference (e.g. 1000 Genomes)

# 3b. Gene expression signature prediction from GWAS summary data using S-PrediXcan

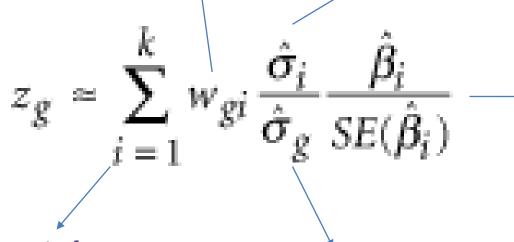


Association of gene *g* expression with phenotype:

+ve z-score:
increased gene
expression
associated with
increased phenotype
or risk of disease

*w<sub>gi</sub>* weight given to each SNP for predicting expression level of *g* Precomputed weights derived from a reference eQTL dataset e.g.
 GTEx

Variance of SNP<sub>i</sub> (from reference genotype data) /



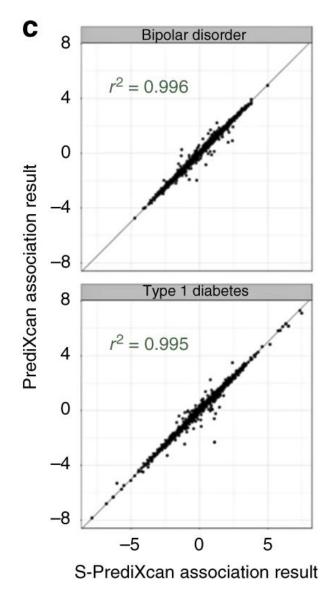
Effect size (and SE) of SNP<sub>i</sub> on phenotype obtained from GWAS

Assuming set of SNP<sub>1..k</sub> contribute to the expression of gene *g* 

Estimated variance of predicted expression of gene *g* 

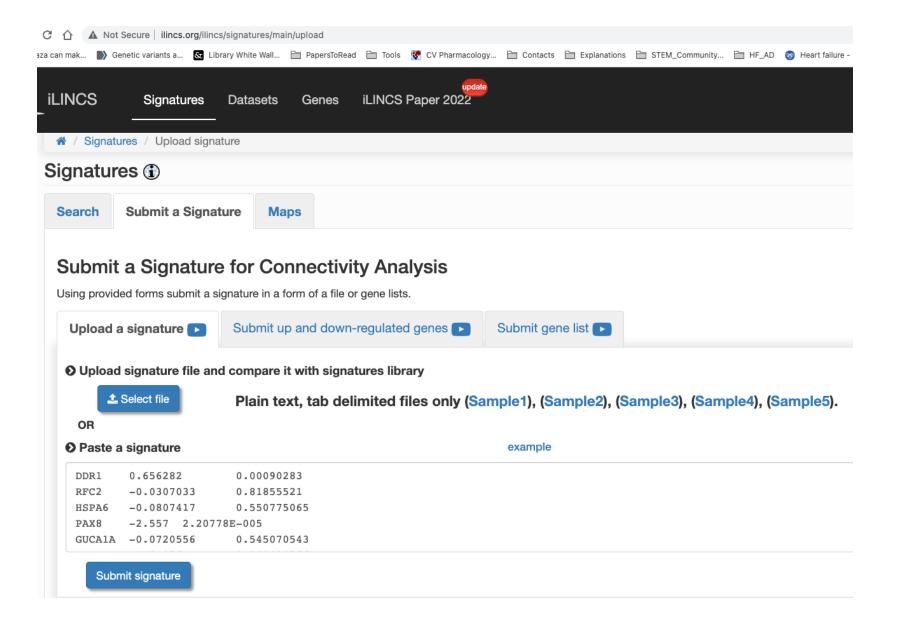
Comparison of PrediXcan and S-PrediXcan gene zscores







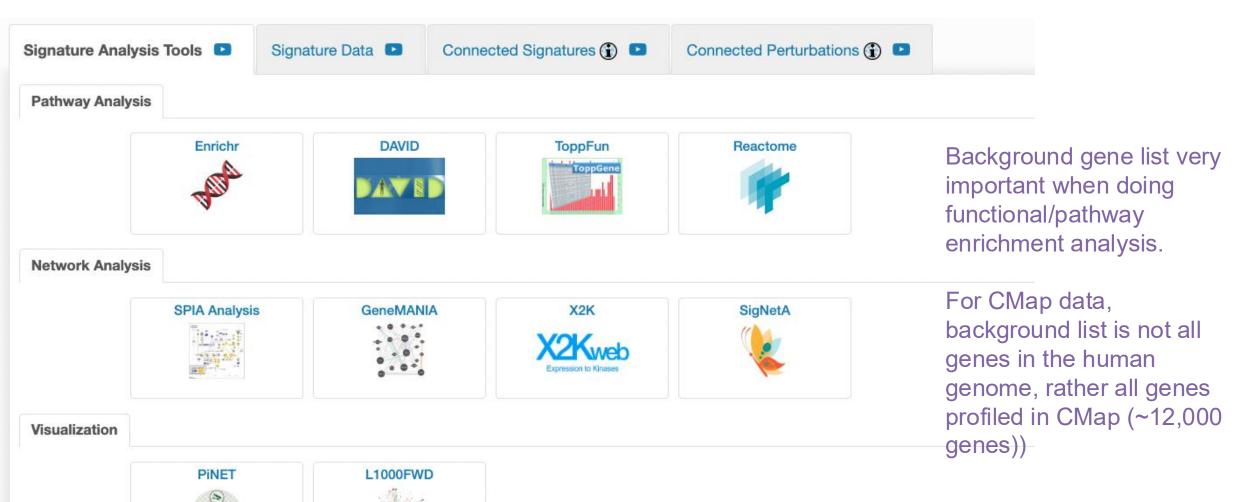
# Querying CMap data with iLINCs http://www.ilincs.org/ilincs/



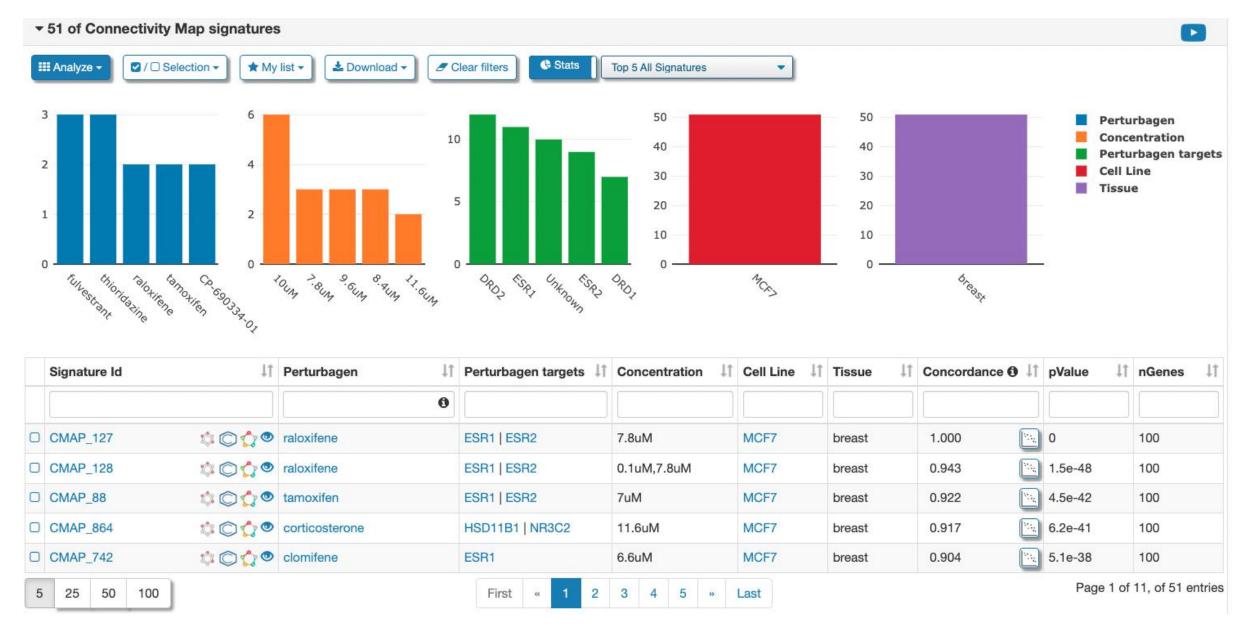










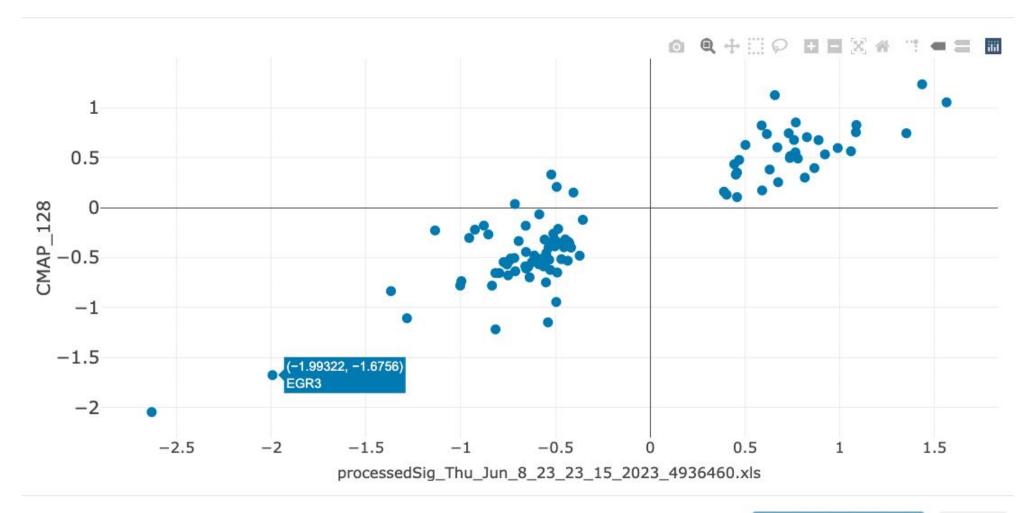




### Correlation plot

Weighted Pearson correlation: 0.943

Pearson correlation: 0.913





### nature neuroscience

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Article | Published: 14 August 2017

# Analysis of genome-wide association data highlights candidates for drug repositioning in psychiatry

Hon-Cheong So <sup>™</sup>, Carlos Kwan-Long Chau, Wan-To Chiu, Kin-Sang Ho, Cho-Pong Lo, Stephanie Ho-Yue Yim & Pak-Chung Sham

Nature Neuroscience 20, 1342–1349 (2017) Cite this article



## So et al

- Brain-based eQTL models to impute disease gene expression
- Spearman, Pearson correlation, and KS-test to determine similarity between between disease and drug signature
- As there are no consensus methods to define K, they set different values of K (50, 100, 250, 500) and averaged the results for each method.
- No selection on cell type



# Important considerations for GWAS to drug candidate pipeline using gene expression signature mapping

- How many genes to include in your query gene list?
- what eQTL model should you use generating a disease gene signature?
  - Single-tissue model (disease tissue-specificity) vs multi-tissue model (greater power to predict gene expression)
- What cell lines should you query in iLINCs?
  - Most relevant cell type vs summary across cell lines
- How does the power of the GWAS affect identification of drug targets?

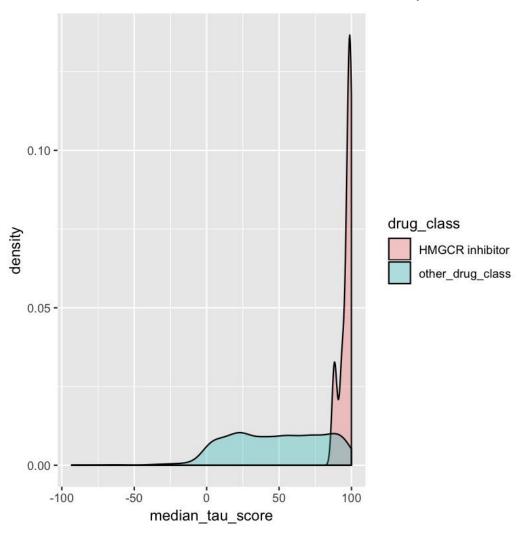


## Other considerations

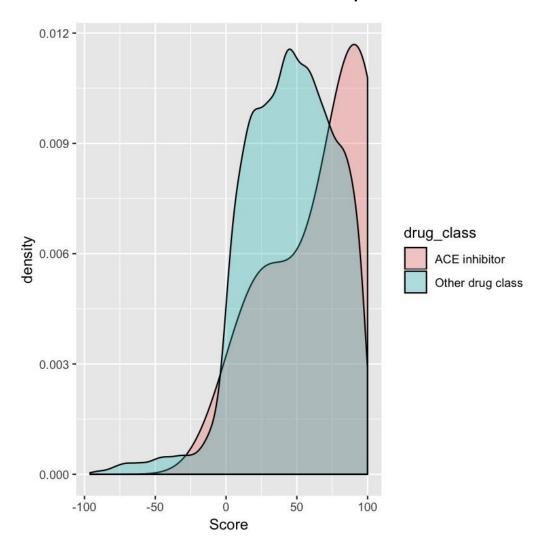
- iLINCS is a useful resource but requires careful manual curation
  - Check connectivity between gene knockdown/overexpression and drug
  - Check specificity of the gene signature
  - Check connectivity between compounds with same MoA
  - Compare genetic vs chemical perturbation signatures (e.g. statin vs HMGCR KO)
  - Check connectivity across cell lines
  - Drugs may not be in an active form. Need to check this from other sources e.g. DrugBank
  - Check if target is expressed in cell line before interpreting results (human protein atlas)



## Connectivity of rosuvastatin with other HMGCR-inhibitors and all other compounds



## Connectivity of enalapril with other ACE inhibitors and all other compounds





### Statins and depression

BMJ. 1992 Feb 15; 304(6824): 431-434.

doi: 10.1136/bmj.304.6824.431

PMCID: PMC188

PMID: 153

Should there be a moratorium on the use of cholesterol lowering drugs?

DEPRESSION

A Statin Island of Woe: Are Corresponder

Depression and Cholesterollowering Connected?

Are statin drugs bringing you down?

2012 Psychology Today



2019 The Times

Do statins have any effects on depression?







Chenwen Hu

Jiayue Clara Jiang

### 1. Connectivity map (CMap) analysis

### 2. Mendelian randomisation analysis

### Translational Psychiatry

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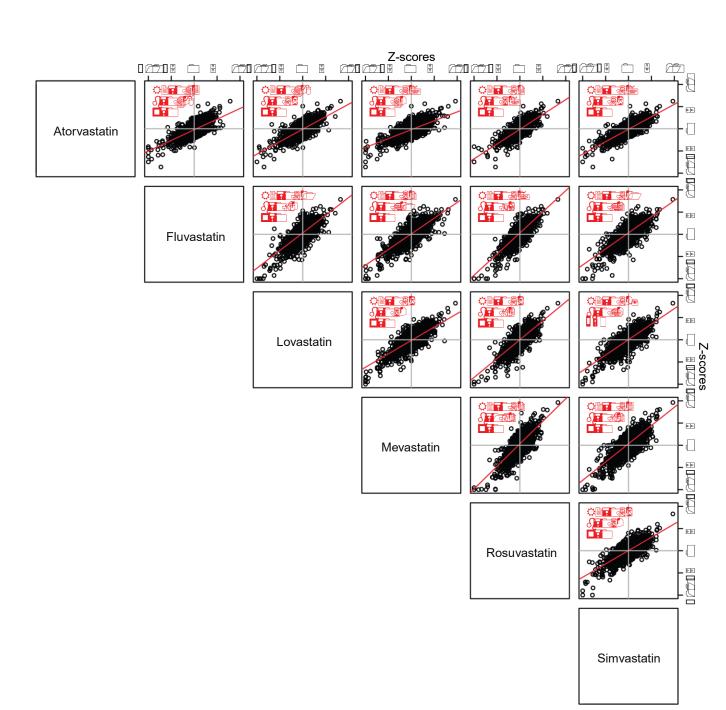
Article Open access | Published: 04 April 2023

# Investigating the potential anti-depressive mechanisms of statins: a transcriptomic and Mendelian randomization analysis

<u>Jiayue-Clara Jiang, Chenwen Hu, Andrew M. McIntosh</u> & <u>Sonia Shah</u> □

<u>Translational Psychiatry</u> **13**, Article number: 110 (2023) <u>Cite this article</u>

**3376** Accesses **3** Citations **14** Altmetric Metrics



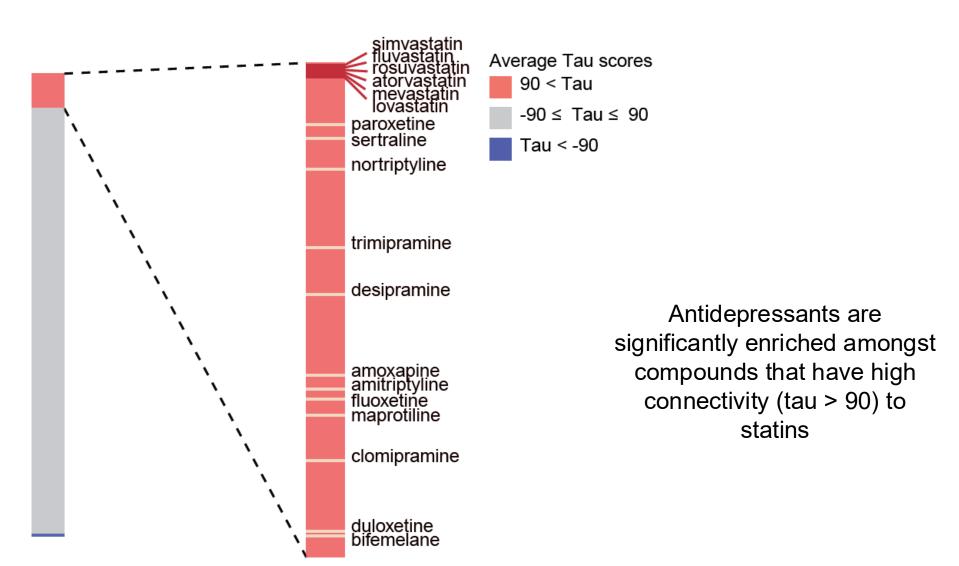


# Within-statin gene expression signature correlation

statin-statin tau scores > 95

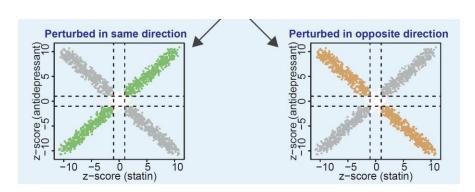


### Enrichment of antidepressants amongst statin-connected compounds

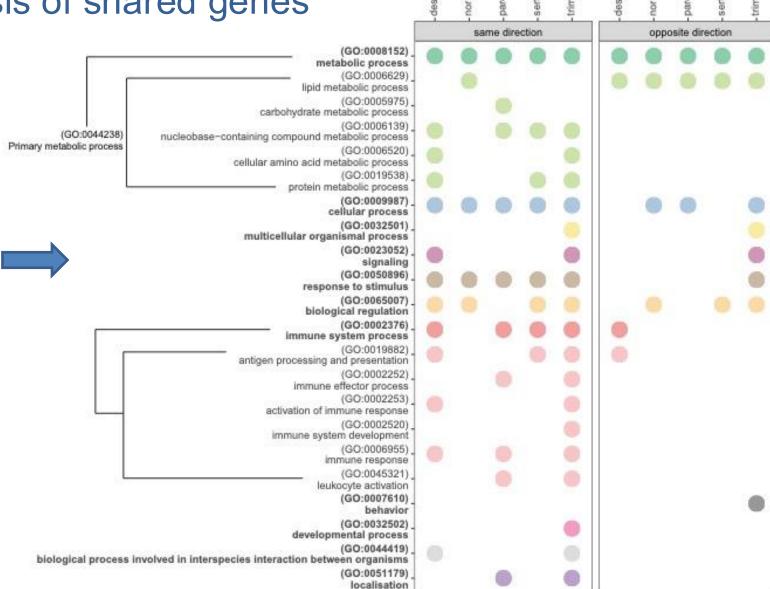




Gene-set enrichment analysis of shared genes

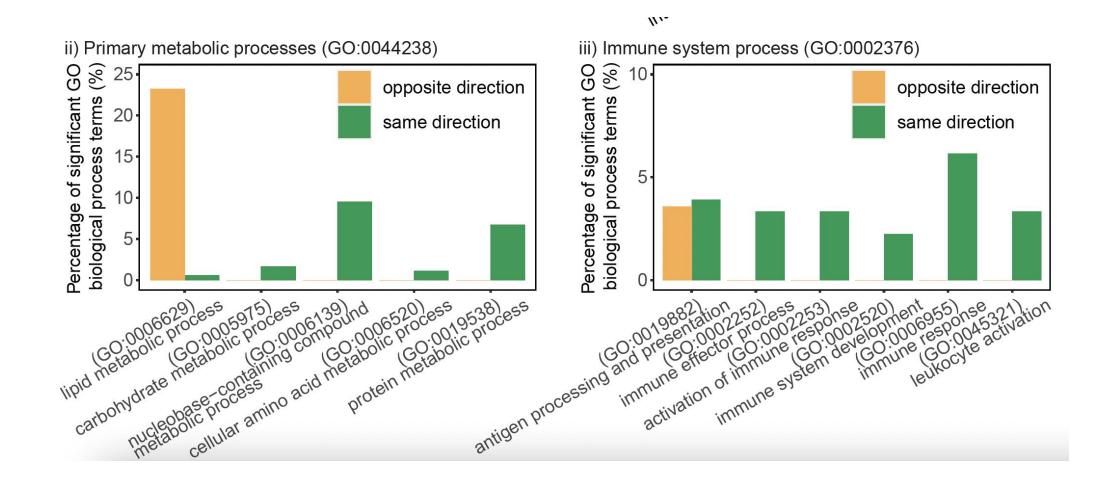


Functional terms enriched amongst genes perturbed in the same and opposite direction by both statins and antidepressants





### Gene-set enrichment analysis of shared genes





# MR analysis HMGCR/ITGAL/HDAC2 gene expression

| Statin       | Gene targets                   |
|--------------|--------------------------------|
| simvastatin  | HMGCR, ITGAL, HDAC2            |
| atorvastatin | HMGCR, DPP4, AHR, HDAC2, NR1I3 |
| rosuvastatin | HMGCR, ITGAL                   |
| lovastatin   | HMGCR, ITGAL, HDAC2            |
| fluvastatin  | HMGCR, HDAC2                   |
| mevastatin   | HMGCR                          |
| cerivastatin | HMGCR                          |
| pitavastatin | HMGCR, ITGAL                   |
| pravastatin  | HMGCR, HDAC2                   |

Drug bank database



Lower HMGCR expression associated with platelet measures

- Significance after multiple testing (SMR p < 0.00057)
- Nominal significance (SMR p < 0.05)

Haematological traits

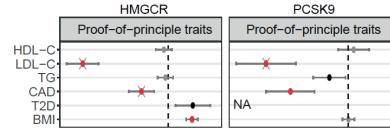
Cytokines

MDD and

related

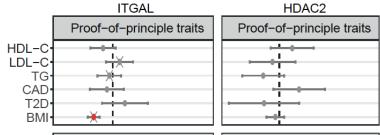
symptoms

- No statistical significance
- X Significance for the HEIDI test (HEIDI p < 0.01)







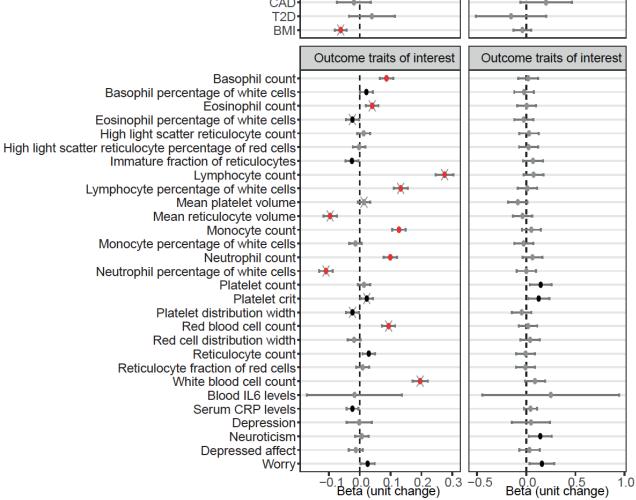


Lower ITGAL expression associated with white blood cell counts



#### Cytokines

MDD and related symptoms





# Your feedback would be greatly appreciated so we can improve on our content next year

- Things you enjoyed and why
- Things you didn't enjoy and why
- Suggestions on how we could improve
- Anything you were hoping we would cover but we didn't?

